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SOIL INOCULATION WITH AZOTOBACTER

BY

PAUL EMERSON

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AGRICULTURAL EXPERIMENT STATION
IOWA STATE COLLEGE OF AGRICULTURE
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AGRONOMY SECTION
Soil Bacteriology

AMES, IOWA

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Soil Inoculation With Azotobacter*

BY PAUL EMERSON.

Following the discovery of the nitrogen fixing powers of the symbiotic bacteria in the soil, early investigators found that the power of utilizing the free atmospheric nitrogen was not confined to the symbiotic bacteria alone. They noted increases in soils which had borne no legumes and they found that fallow soils in particular increased appreciably in nitrogen content. These facts stimulated researches which led to the discovery of many forms of bacteria which are able, when growing alone, to fix nitrogen from the air. The chief of these is now known as the azotobacter group.

It seems likely that the azotobacter will prove more effective in fixing nitrogen than the symbiotic bacteria, although the general requirements of the two classes of organisms are very similar. The azotobacter are active in practically all soils regardless of the kind of crop grown when conditions for their growth are satisfactory. These conditions are probably much the same as for the symbiotic bacteria except that these latter organisms require the presence of a specific legume for fixing the greatest amount of nitrogen. Azotobacter require a certain amount of carbonaceous material in the soils and are usually stimulated by a small amount of nitrogen, but the exact optimum conditions for their growth are as yet unknown. These organisms are active in causing nitrogen increases in many soils, but the feasibility of introducing them into the soil or of attempting to increase their nitrogen-fixing powers by artificial means, and the effect of the presence of growing plants on their efficiency are questions as yet unanswered, although Lipman has indicated that under proper conditions successful inoculation may be accomplished in soils and Bottomley has successfully grown pure cultures of these organisms in the presence of growing plants with favorable results.

HISTORICAL

Beijerinck (2) isolated and described the first azotobacter (in 1901). He found two species, one of which he named *Azotobacter chroöcoccum* and the other *Azotobacter agilis.* The former was isolated from the soil and the latter from a sample of water taken from one of the canals in the city of Delft. Two years later Lipman (36) added a third species, *A. vinelandii,* to the list and the following year isolated and described two more, giving them the names of *A. beijerinckii* and *A. woodstownii.* Of the five organisms of this species, *A. chroöcoccum, A. beijerinckii*

*Thesis submitted in partial fulfillment of the requirements for the Degree of Doctor of Philosophy at the Iowa State College.

and *A. vinelandii* are considered the most important in soil inoculation studies.

The frequency with which investigators in all parts of Europe and America have isolated azotobacter from various soils, indicates that they are widely distributed. Christensen (10) found that they were present throughout northwestern Europe, the activity of the organism apparently depending on the basicity of the soil. This view was later supported by the works of Fisher (14), Löhnis and Pillai (45) and others. Ashby (1) studied the soils of Mombasa, East Africa, Cairo, Egypt and Rothamsted, England and found azotobacter forms present in most cases. Lipman and Burgess (42) working with forty-six samples of soil from various parts of the world, found that over one-third of them contained azotobacter, the predominant form being *A. chroöcoccum*. Many of the soils examined were museum specimens and had been kept in tightly stoppered bottles for long periods of time.

DESCRIPTION OF AZOTOBACTER.

Beijerinck characterizes the azotobacter as stout bacteria, 4-6 microns or less in length, sometimes longer, occurring as large diplococci or short rods in young cultures, the hyaline cells often containing a vacuole and the entire organism enclosed in a mucilagenous wall of varying thickness. They have a single polar flagellum or bundles of 4-10 polar flagella of about the same length as the organism itself. Beijerinck found no spores. Vagler (65) writes that the older colonies produce involution forms similar to those of yeasts while Heinz (22) and Fisher (15) showed that the organisms can resist drying for six to nine months. Later investigations by Mulvania (50) and Löhnis and Smith (47) demonstrated that the organism produces spores and completes a very complicated life cycle. Descriptions of azotobacter and detailed cultural characteristics of the organism were given by Lipman (35), Beijerinck (2), Prazmowski (54), Warmbold (70), Bonazzi (6), Löhnis and Westerman (48), Löhnis and Hanzawa (44), Jones (27) and others.

ACTIVITIES OF AZOTOBACTER.

Beijerinck first claimed that the isolated pure cultures of azotobacter were able to fix the atmospheric nitrogen in appreciable amounts; later, however, when working with Van Delden (4), he retracted this statement, claiming that pure cultures did not have this ability and that only in the presence of very small celled organisms called radiobacter could the free nitrogen of the air be fixed in the soil. Gerlach and Vogel (18), Heinz (23), Lipman (37) and Freudenreich (17) proved conclusively that the earlier conclusions of Beijerinck were correct and that

the organism may fix considerable amounts of nitrogen in pure cultures. Lipman accounts for the fact that Beijerinck did not get a fixation of nitrogen in pure cultures by showing that the organism will not fix nitrogen unless the reaction of the medium is made neutral or slightly alkaline. When Beijerinck later accepted this suggestion he found that his pure cultures were able to fix atmospheric nitrogen.

STUDIES OF AZOTOBACTER.

Very few investigators have attempted to inoculate soils with azotobacter or other non-symbiotic nitrogen fixing bacteria under conditions approximating those in the field. The influence of various kinds of sugars, cellulose, inorganic salts, and various organic compounds on the nitrogen-fixing power of the organisms have been studied extensively. Gerlach and Vogel (19), Pringsheim (55), Krainsky (33), Koch (30), Hoffman and Hammer (25) and Stranak (61) have found that various sugars and cellulose materially increase their nitrogen fixing powers while Fisher (16), Christensen (10), Löhnis and Pillai (46), Wilfarth and Wimmer (59) Kaserer (28), Rosing (59), Vogel (66), Greaves and Anderson (20) and Pringsheim (56) have shown that small amounts of lime, very small amounts of nitrogen, various inorganic salts and even a very small amount of arsenic will stimulate the nitrogen fixing power of the organisms in the presence of certain carbon compounds. Stoklasa (60) studied the products of the activities of the azotobacter organisms, confining his researches largely to the amounts and kinds of gases produced under different circumstances, under the influence of various substances supposed to be energy sources, and under varying temperature conditions. His results have been more or less confirmed by the works of Thiele (64), Hoffman (24), Kellerman and Smith (29) and Ehrenberg (13).

The activity of the azotobacter in soils in general, and particularly under laboratory conditions, was fully shown by the works of Lipman (39), Voorhees and Lipman (68), Löhnis (43), Kuhn (34), Freudenreich (17), Dvorak (12), Remy (57), Remy and Rosing (58), Jacobitz (26), Stranak (62), Headden (21), Peterson and Mohr (52), Koch and Seydel (31), Omeliansky and Ssewerowa (51), Warmbold (71) and others who demonstrated that under various conditions and in almost every type of known soil these organisms are able to fix appreciable amounts of the free atmospheric nitrogen. Only a few of these investigators, however, have made any attempt to secure an active flora of these organisms in the soil. Vogel (67) inoculated pure cultures of azotobacter into soils that had been treated with grape sugar, in some cases adding comparatively large amounts of nitrate of soda. In pot experiments with oats and mustard, increases were

noted for the inoculated series, altho the pots receiving nitrate of soda gave the greatest yields. When the experiment was repeated in the field the inoculated plots gave smaller yields than the uninoculated, and the inoculation appeared to have an injurious effect upon the crop.

A short time later Lipman and Brown (41) tried inoculation experiments with *A. vinelandii* and *A. beijerinckii.* They sunk four foot cylinders open at both ends into the soil, filled the cylinders with soil and inoculated the soil with the organisms. The first summer the soils were left bare and then a rotation of crops was followed and oats, corn and rye grown in succession. While considerable variation was found in the nitrogen content of the crops and in the dry weight, the general conclusion reached was that the activities of the organisms did not increase the nitrogen content of the soil. The results do not preclude the possibility that inoculation with the organisms in question may be made of practical value, provided proper conditions for the best growth of the organisms are secured. Bottomley (7) and Bottomley and Hall (9) experimented with oats, barley and some root crops, and arrived at the same conclusions as did Lipman and Brown. Stranak (63) also inoculated soils with azotobacter and found a pronounced increase in the growth of potatoes, grain and beets.

Altho the experiments dealing with the inoculation of soils with azotobacter have been inconclusive, it is believed that under proper conditions such inoculation may be extremely profitable.

EXPERIMENTAL

The wide distribution of non-symbiotic nitrogen fixing bacteria in many types of soils is practically parallel with the distribution of the symbiotic organisms, and since it is practical and profitable to inoculate soil with the latter, even tho the particular organism may be present, the following questions quite naturally arise:

1. If the azotobacter are not present in the soil, can inoculation be profitably accomplished?
2. What soil conditions are necessary for the greatest fixation of nitrogen by these organisms?

These questions have an important bearing on the problem of the maintenance of permanent fertility in soils from the nitrogen standpoint and may govern the choice of the proper method of farming. Some commercial concerns have placed cultures on the market, claiming that they contain sufficient numbers of the non-symbiotic nitrogen fixing bacteria to enable the farmer to solve his nitrogen problem without growing legumes. However, results of experiments showing that such cultures are capable of inoculating the soil were not found in the present investigation.

INCREASING THE NITROGEN FIXING POWER OF PURE CULTURES.

Very little work has been done along the line of breeding pure cultures of bacteria to an increased efficiency in their specific actions, in fact, practically all the experiments have been carried out with the idea of finding a method whereby the organism could be kept alive for long periods without periodic transfers. The earliest investigation along this line was that of Czaplewski (11) who limited the amount of air in the tube by saturating the plug with paraffine. Later Lunt (49) found that certain cultures of water bacteria may be kept alive much longer in sterile water than in ordinary culture media. In some cases he kept certain organisms alive for two years by this method. Bolley (5) secured good growths of *B. amylovorus* and *Bact. dianthi* in agar and in bouillon by making transfers from cultures that had been hermetically sealed for nine years. It is not stated whether or not the organisms were tested for their pathogenicity and hence their virulence is left in doubt. This work supports that of Czaplewski in showing that cultures can be kept alive for long periods of time if the transpiration is reduced to a minimum. Some commercial concerns claim that they are able to increase the efficiency of their particular cultures of legume bacteria by alternate inoculations first on agar, then into sterile greenhouse soil, growing the specific legume to which the organism in question is adapted, and re-isolating the organism from the nodules produced on the roots of the legume. If this is possible for the symbiotic bacteria then it seems probable in the case of the non-symbiotic organisms. The following questions naturally suggest themselves:

1. Can the nitrogen fixing power of azotobacter be increased by periodic transfers on nitrogen free media?
2. Can the nitrogen fixing powers of azotobacter be increased by growing the organism in the presence of growing plants?

In outlining work to answer the above questions it was realized that a large number of bacteria should be used. A number of large celled nitrogen fixing organisms that had all the staining reactions of the azotobacter type and closely resembled it in size and shape, were isolated in pure cultures from soil secured from the humus plots at the Iowa station and were designated with laboratory numbers. At the same time pure cultures were secured and their activities determined along with those of the unnamed cultures. The pure cultures were kindly furnished by Dr. J. G. Lipman of the New Jersey Agricultural Experiment Station and also by the American Museum of Natural History of New York.

MEDIA USED.

The nitrogen free medium used thruout the experimental work was a modification of that proposed and used by Lipman (35), and its composition was as follows:

Distilled water	1,000 cc
Di-potassium phosphate	0.2 grams
Magnesium sulphate	0.2 grams
Calcium chloride	0.02 grams
Dextrose	10.0 grams
10% Ferric chloride solution	2 drops

The solution was brought to boiling and made neutral to phenolphthalein by the addition of N/10 NaOH. If a solid medium was desired 1% powdered agar was added. Sterilization was accomplished by placing in the autoclave at ten pounds for 20 minutes.

Inoculation was secured by scraping off a two days' growth from the agar slants with a sterile needle and transferring it to flasks containing 50 cc. of the above solution. In order to determine whether the nitrogen fixing power of the organisms was stimulated by the addition of nitrogen, the above solution with the addition of 1 mg. of nitrogen as sodium nitrate was used.

PRELIMINARY STUDIES.

All of the organisms of the azotobacter type including both the pure cultures and the unnamed cultures, were inoculated into 50 c. c. of both of the above solutions and tested for their nitrogen fixing powers. The inoculated solutions were incubated for three weeks at room temperature (22-25° C) and then Kjeldahlized. The amount of nitrogen fixed by each organism in the different solutions is shown in table I. The same cultures were transferred 12 times at three to four day intervals on nitrogen free media and their nitrogen fixing power tested in solutions with and without nitrogen. The results appear in table II.

The laboratory organisms used in table I had been freshly isolated and purified from the soil, the named cultures had been kept on agar slants for varying periods of time. During the time that the inoculated culture solutions were incubating the transfers were being made in preparation for the inoculations for table II.

Comparing the two tables we find that a majority of the organisms decreased in their ability to fix atmospheric nitrogen, altho a few showed a slight increase or at least retained their efficiency. From these the following eight were selected for further study: No. 4, No. 22, No. 26, No 27, *A. vinelandii*, *A. chroococcum*, *A. beijerinckii* and *A. chroococcum* (HCM). These eight organisms were studied under both laboratory and greenhouse conditions.

TABLE I—NITROGEN-FIXATION BY PURE CULTURES IN SOLUTION WITH AND WITHOUT NITROGEN.

Organism	N. Fixed in Mgs.	
	Solution without Nitrogen	Solution with Nitrogen
Lab. No. 1	2.24	0.14
Lab. No. 2	2.38	1.54
Lab. No. 3	0.28	----
Lab. No. 4	0.98	1.96
Lab. No. 6	1.96	1.82
Lab. No. 7	3.22	1.68
Lab. No. 9	0.42	2.10
Lab. No. 10	0.42	0.84
Lab. No. 11	0.42	0.70
Lab. No. 12	lost	1.40
Lab. No. 14	1.12	lost
Lab. No. 15	2.80	1.12
Lab. No. 16	7.14	lost
Lab. No. 18	0.28	0.42
Lab. No. 19	1.12	1.54
Lab. No. 20	0.56	1.96
Lab. No. 21	0.56	2.10
Lab. No. 22	0.42	2.52
Lab. No. 23	0.70	2.52
Lab. No. 24	0.70	1.82
Lab. No. 25	0.28	1.82
Lab. No. 26	0.56	2.10
Lab. No. 27	1.12	5.60
A. vinelandii	lost	2.66
A. chroococcum (HCM)	4.20	3.08
A. chroococcum	0.84	1.54
A. chroococcum (Colo)	0.70	2.52
A. beijerinckii	0.84	1.68
A. beijerinckii No. 5	1.96	2.38

TABLE II—NITROGEN-FIXATION BY PURE CULTURES IN SOLUTION WITH AND WITHOUT NITROGEN.

After each organism has been transferred twelve times on nitrogen-free media at three to four day intervals.

Organism	N. Fixed in Mgs.	
	Solution without Nitrogen	Solution with Nitrogen
Lab. No. 1	0.84	1.12
Lab. No. 2	0.14	0.00
Lab. No. 3	0.14	0.98
Lab. No. 4	1.82	0.98
Lab. No. 6	0.00	0.84
Lab. No. 7	0.28	0.28
Lab. No. 9	0.00	1.26
Lab. No. 10	0.98	0.00
Lab. No. 11	0.00	1.68
Lab. No. 12	0.00	0.28
Lab. No. 14	0.42	1.12
Lab. No. 15	0.00	1.40
Lab. No. 16	0.00	0.14
Lab. No. 18	0.00	0.42
Lab. No. 19	0.00	0.42
Lab. No. 20	0.98	1.54
Lab. No. 21	0.42	0.84
Lab. No. 22	2.52	1.12
Lab. No. 23	2.10	1.54
Lab. No. 24	0.00	1.68
Lab. No. 25	0.00	0.28
Lab. No. 26	1.12	1.82
Lab. No. 27	1.40	2.52
A. vinelandii	0.00	0.00
A. chroococcum	1.12	1.82
A. chroococcum (HCM)	2.52	1.12
A. chroococcum (Colo)	0.00	1.26
A. beijerinckii	0.42	2.52
A. beijerinckii No. 5	1.12	0.00

LABORATORY STUDIES.

The laboratory studies were arranged in a series of three experiments as follows:

1. To determine the effect of transfers made every other day on the nitrogen fixing power of the organisms.

2. To determine the effect of transfers made once each week in sand cultures variously modified.

3. To determine the effect of growing four of the organisms on both agar and sand in large flasks with and without the presence of growing plants.

Series 1. To Determine the Effect of Transfers Made Every Other Day on the Nitrogen Fixing Power of the Organisms.

Using the eight selected organisms transfers were made every other day on the nitrogen free medium for a period of three weeks. It was feared that such rapid transferring for so long a period on a medium practically free from nitrogen would reduce the vitality of the organisms, accordingly each fifth transfer was made on a modification of the medium consisting in the addition of one milligram of nitrogen as sodium nitrate to each liter of the regular dextrose agar. At the end of the transfer period the organisms were inoculated into the nitrogen free and nitrogen containing solutions incubated for the same periods of time and the amount of nitrogen fixed determined by Kjeldahlizing. The results of the determinations are shown in table III.

TABLE III—THE EFFECT OF TRANSFERS MADE EVERY OTHER DAY FOR FOUR WEEKS ON THE NITROGEN FIXING POWER OF THE ORGANISMS.

Organism	Nitrogen Fixed in Mgs.					
	Solution without Nitrogen			Solution with Nitrogen		
	(a)	(b)	(Av.)	(a)	(b)	(Av')
Lab. No. 4	0.14	0.42	0.28	0.70	0.42	0.56
Lab. No. 22	0.00	0.00	0.00	0.14	0.56	0.35
Lab. No. 26	0.14	0.14	0.14	0.28	0.98	0.63
Lab. No. 27	0.28	3.50	1.98	0.70	0.98	0.84
A. vinelandii	0.28	0.42	0.35	0.98	0.98	0.98
A. chroococcum	0 56	0.28	0.42	0.98	1.12	1.05
A. chroococcum (HCM)	2.66	lost	2.66	1.40	1.40	1.40
A. beijerinckii	0.84	2.66	1.75	0.28	0.28	0.28

That these transfers should have been made at longer intervals is evidenced by the fact that tables I and II showed that 12 of the cultures had increased in efficiency after they had been transferred every three days for 36 days. However, during the latter work the organisms did not show any indications of a loss of vitality and the growth at all times was vigorous and rapid. Table III shows a decrease in the nitrogen fixing powers of all the organisms except in the case of A. chroococcum (HCM) which appears to have retained its efficiency thruout the experiment.

Series 2. To Determine the Effect on the Nitrogen Fixing Power of Transfers Made Each Week in Sand Cultures.

In the following experiment sand was used instead of agar as the basis for the medium. Ground oats straw, ground red clover hay and either the regular dextrose solution, or the dextrose solution containing nitrogen were added. The tests were carried out in tubes arranged as follows:

6.25 gr. sand+2.5 cc N. free dextrose solution.
6.25 gr. sand+2.5 cc dextrose solution containing 0.2 gr. $NaNO_3$ per liter.
6.25 gr. sand+3.5 cc N. free dextrose solution+0.1 gr. clover hay.
6.25 gr. sand+3.5 cc N. free dextrose solution+0.1 gr. oats straw.

The organisms were transferred directly from the slants into the tubes and there allowed to incubate at room temperature for seven days. A small portion of the sand was then transferred to a fresh tube of the same medium as the original. As this particular experiment did not directly follow the others the efficiency of the organisms was tested before they were inoculated into the sand. Table IV shows the amount of nitrogen fixed by the pure

TABLE IV—THE NITROGEN FIXING POWER OF THE PURE CULTURES IMMEDIATELY BEFORE THE SAND CULTURE EXPERIMENTS.

Organism	[Solution witnout Nitrogen			Solution with Nitrogen		
	(a)	(b)	(Av.)	(a)	(b)	(Av.
Lab. No. 4	0.14	0.70	0.42	0.28	1.54	0.91
Lab. No. 22	0.00	0.00	0.00	3.36	3.08	3.22
Lab. No. 26	0.00	0.14	0.07	2.94	2.66	2.80
Lab. No. 27	0.00	0.00	0.00	2.38	2.52	2.45
A. vinelandii	0.00	0.00	0.00	1.40	1.40	1.40
A. chroococcum	0.00	0.14	0.07	2.80	2.66	2.73
A. chroococcum (HCM)	0.00	0.00	0.00	2.52	2.38	2.45
A. beijerinckii	0.98	0.84	0.91	2.94	lost	2.94

cultures at the beginning of this series of incubation, and the same methods as in the previous experiments.

At the end of the fourth transfer period, i. e., four weeks after the start of the experimental series, the organisms were inoculated into dextrose solution and their nitrogen fixing powers determined. After four more weeks of transferring or in all eight weeks the final inoculation into dextrose solution was made. The influence of the oats and clover in the presence of sand on the nitrogen fixing power of the organisms used is shown in tables V and VI, by the fact that both the large celled organisms of the

TABLE V.—NITROGEN FIXED BY THE PURE CULTURES AFTER FOUR TRANSFERS IN SAND AT PERIODS OF SEVEN DAYS EACH.

Organism	Nitrogen Fixed in Mgs.			
	dex. sol.	dex. sol. + N	dex. sol.+ oats straw	dex. sol.+ clover hay
Lab. No. 4	0.28	0.35	0.28	0.98
Lab. No. 22	0.07	0.28	0.00	0.35
Lab. No. 26	0.77	0.07	0.14	0.00
Lab. No. 27	0.42	0.35	1.27	0.42
A. vinelandii	0.14	0.21	0.42	0.28
A. chroococcum	0.21	0.21	0.07	0.21
A. chroococcum (HCM)	0.07	0.14	1.19	0.42
A. beijerinckii	0.14	0.28	0.35	0.28

TABLE VI—NITROGEN FIXED BY THE PURE CULTURES AFTER EIGHT TRANSFERS AT PERIODS OF SEVEN DAYS EACH.

Organism	Nitrogen Fixed in Mgs.			
	dex. sol.	dex. sol. + N	dex. sol.+ oats straw	dex. sol.+ clover hay
Lab. No. 4	0.30	0.20	0.30	0.40
Lab. No. 22	0.20	0.20	0.20	2.00
Lab. No. 26	0.70	0.40	0.00	1.00
Lab. No. 27	0.20	0.10	0.40	0.50
A. vinelandii	0.20	0.40	0.40	0.50
A. chroococcum	0.30	0.10	0.20	lost
A. chroococcum (HCM)	0.10	0.40	0.40	1.40
A. beijerinckii	0.00	0.00	0.20	0.50

azotobacter type and the azotobacter themselves, made gains in their nitrogen fixing powers. There was no distinct gain due to any one kind of carbonaceous material. Of the six organisms showing gains *A. chroococcum* made the most notable, especially in the presence of the oats straw. The nitrogen fixing power of No. 4 appears to be rather constant thruout the series, with no appreciable gain or loss. *A. beijerinckii* showed a decided loss in its power to fix nitrogen in each of the four media, but gave a slight indication that in the presence of the clover hay it might be slowly regaining its power.

Series 3. To Determine the Effect of Growing the Organisms on Both Agar and Sand With and Without the Presence of Growing Plants.

The main points considered in this experiment were: An increase in the surface area over which the organism could grow; an increase in the time between transfers and the growing of the organisms in the presence of an undetermined species of algæ and with growing oats and red clover plants. Two liter Erlenmeyer flasks were used and arranged in the following manner conforming to the outlines of the experiment:

Flask No. 1. 1000 cc N. free dex. agar+1 gr. $CaCO_3$ planted to oats.

Flask No. 2. 1000 cc N. free dex. agar+1 gr. $CaCO_3$ planted to red clover.

Flask No. 3. 1000 cc N. free dex. agar+1 gr. $CaCO_3$ planted with an undetermined species of algæ.

Flask No. 4. 1000 gr. pure quartz sand+180 cc N. free dex. solution neutralized with $CaCO_3$, planted with oats.

Flask No. 5. 1000 gr. pure quartz sand+180 cc N. free dex. solution neutralized with $CaCO_3$, planted with red clover.

Flask No. 6. 1000 gr. pure quartz sand+180 cc solution without dex. neutralized with $CaCO_3$, planted with an undetermined species of algæ.

Check flasks of sand and dextrose agar.

The flasks of agar were sterilized in the autoclave at ten pounds for 30 minutes, but the flasks of sand were sterilized at 15 pounds pressure for four hours once a day for three consecutive days. Bacteriological tests on the sand at the end of that time showed it to be sterile.

The culture of the algae used was so closely associated with a bacterial growth that a separation would have required a long time. For that reason it was not purified but was grown in sterile distilled water, for about three months before inoculation. The inoculation of the algae was made in the flasks about two weeks before the inoculation with the azotobacter cultures in order that the algae might make a sufficient growth to supply the bacterial cultures with the proper amount of carbonaceous material. To prevent contamination by the oat and red clover plants, the seeds were soaked three minutes in a 1-500 solution of mercuric chloride, washed in sterile distilled water three times and then planted in sterile agar plates. By this means the seeds were sprouted and those which were contaminated were discovered and rejected. The sprouted seeds were transferred from the plates to the flasks by means of the platinum needle. A block of the agar containing the sprouted seed was cut out and placed in the proper position on the medium in the flask. The flasks were then carefully observed for five days to insure the absence of contamination.

As all the flasks contained growing plants no attempt was made to exclude the light, but neither were they placed in the direct sunlight. They were kept on a table about eight feet from a large window facing the west. All the flasks were plugged with non-absorbent cotton and after inoculation a cap of paraffined paper was placed over the mouth and held in place with a rubber band. While the plants did not develop rapidly the oats grew much faster than the clover for about three weeks, after which time both began to lose chlorophyl and by the end of the five weeks' experimental period, the majority of the plants had died. The oats and clover in the flasks inoculated with A. chroococcum (HCM) and the clover in the flasks inoculated with B. radicicola showed a slight gain in growth and altho far from vigorous at the end of the experiment were still alive and growing slowly.

ORGANISMS USED.

The organisms used were *A. chroococcum* (HCM), *A. vinelandii*, *A. beijerinckii* and for the purpose of comparison, *B. radicicola* isolated from the nodules of sweet clover. The latter were isolated and purified especially for this series, and introduced to compare the effects of symbiotic and non-symbiotic organisms on the growth of the plant used. The results secured with it, however, were of no great significance.

After the bacteria had remained undisturbed in the flasks for five weeks, transfers were made directly from the flasks into 50 cc. of the nitrogen-free dextrose solution, incubated for three weeks, and the nitrogen fixed determined in the usual manner. The total amount of nitrogen fixed by the bacteria themselves, as well as the amount fixed by the bacteria but due to the stimulative action of the plants on the bacterial activities, is shown in table VII. There was a stimulation of the nitrogen fixing power of the organisms due to the presence of a growing plant, especially noticeable in the case of *A. vinelandii* and *A. chroococcum* (HCM) and to some extent in the case of *A. beijerinckii*. *A. vinelandii* was stimulated thruout the entire series except when grown in sand in the presence of the algae. The oats and algae showed no

TABLE VII—THE EFFECT OF GROWING PLANTS ON THE NITROGEN FIXING POWER OF PURE CULTURES.

Inoculum	Medium	Plant Used	Nitrogen Fixed in Mgs. (a)	(b)	(Av)	N. in algae check	Total amount due to bact. action.	Nitrogen in sand or agar check.	Total amount due to stimulative action of plants.
Algae	agar	check	0.84	0.84	0.84				
Algae	sand	check	1.40	0.98	1.19				
Vinelandii	agar	check	1.12	0.98	1.05				
Vinelandii	sand	check	1.40	1.40	1.40				
Vinelandii	agar	oats	3.66	3.52	3.59		3.59	1.05	2.54
Vinelandii	agar	red clover	2.10	2.38	2.24		2.24	1.05	1.19
Vinelandii	agar	algae	4.20	lost	4.20	0.84	3.36	1.05	2.31
Vinelandii	sand	oats	4.20	4.06	4.13		4.13	1.40	2.73
Vinelandii	sand	red clover	4.48	4.06	4.27		4.27	1.40	2.87
Vinelandii	sand	algae	2.52	2.80	2.66	1.19	1.47	1.40	0.07
Chroococcum (HCM)	agar	check	0.00	0.00	0.00				
Chroococcum (HCM)	sand	check	0.28	0.14	0.21				
Chroococcum (HCM)	agar	oats	0.28	0.44	0.36		0.36	0.00	0.35
Chroococcum (HCM)	agar	red clover	1.82	3.22	2.52		2.52	0.00	2.52
Chroococcum (HCM)	agar	algae	3.92	3.50	3.71	0.84	2.87	0.00	2.87
Chroococcum (HCM)	sand	oats	0.28	0.56	0.42		0.42	0.21	0.21
Chroococcum (HCM)	sand	red clover	0.28	lost	0.28		0.28	0.21	0.07
Chroococcum (HCM)	sand	algae	3.22	5.18	4.20	1.19	3.01	0.21	2.80
Beijerinckii	agar	check	0.00	0.00	0.00				
Beijerinckii	sand	check	0.28	0.42	0.35				
Beijerinckii	agar	oats	0.14	0.00	0.07		0.07	0.00	0.07
Beijerinckii	agar	red clover	1.40	1.40	1.40		1.40	0.00	1.40
Beijerinckii	agar	algae	0.00	0.00	0.00	0.84	-----	-----	-----
Beijerinckii	sand	oats	1.39	1.39	1.39		1.39	0.35	1.04
Beijerinckii	sand	red clover	1.39	1.39	1.39		1.39	0.35	1.04
Beijerinckii	sand	algae	1.39	1.40	1.40	1.19	0.21	0.35	-----
B. rad., S. clo.	agar	oats	0.14	0.14	.014		0.14		
B. rad., S. clo.	agar	red clover	0.14	0.14	.014		0.14		
B. rad., S. clo.	agar	algae	0.56	0.56	0.56	0.84	-----		
B. rad., S. clo.	sand	oats	0.42	0.00	0.21		0.21		
B. rad., S. clo.	sand	red clover	0.14	0.00	0.07		0.07		
B. rad., S. clo.	sand	algae	0.42	0.56	0.49	1.19	-----		

difference when grown on the agar and in the sand medium the greatest stimulation was produced by the red clover. The activities of *A. chroococcum* were stimulated to the greatest extent by the presence of algae in both sand and agar, the oats gave a poor stimulation in both cases, and red clover gave good results in the agar but not in the sand.

The nitrogen fixing power of *A. beijerinckii* was retarded by the presence of algae, but was stimulated by red clover in both the agar and sand. The oats stimulated this organism only when grown on the agar. The nitrogen fixing power of *B. radicicola* was so low thruout the experiment that the results are not considered.

CONCLUSIONS FROM LABORATORY STUDIES.

1. Transfers made on a nitrogen free dextrose agar more often than once each week were detrimental to the nitrogen fixing power of azotobacter and other large celled nitrogen fixing organisms of the same type.

2. Transfers made once each week into a pure sand medium containing some carbonaceous material were beneficial to the nitrogen fixing power of the azotobacter in general, but the effect on *A. beijerinckii* was detrimental.

3. The nitrogen fixing power of *A. vinelandii* was stimulated to a marked extent when grown in large flasks for five weeks in the presence of red clover and oats on both agar and sand. It was stimulated by the presence of algae when grown on agar but not when grown on sand.

The nitrogen fixing power of *A. chroococcum* was stimulated markedly when grown on agar for five weeks in the presence of growing oats and red clover, but to a less extent when grown with the same plants in sand. The greatest stimulation for this organism was produced by growing it in the presence of algae in either sand or agar for the same period of time.

5. The nitrogen fixing power of *A. beijerinckii* was stimulated by the presence of red clover when grown on either sand or agar, and by oats when grown in sand. Algae in either agar or sand appeared to have a depressing effect on the nitrogen fixing power of this organism.

GREENHOUSE STUDIES.

At the conclusion of the first experiment the eight organisms used in the laboratory series 1, 2 and 3 were also inoculated into soils in pots in the greenhouse. Ground oats straw or ground clover hay was added to these soils and the nitrogen fixing efficiency of the organisms both in fallow soils and in the presence of growing oats plants determined. Three experiments were carried out in this test, as soon as the soil in which one crop had been

grown was sampled, it was immediately reseeded and another crop grown. Strict account was kept of the amount of nitrogen added in the seed and in the organic matter. The dry weight of the crop and the N. content as well as the nitrogen content of the soil was determined at the end of each experiment.

The soil used thruout the experiment was of the type classified by the United States Bureau of Soils as Miami silt loam, and according to tests in the laboratory did not contain azotobacter or any similar organisms. A large amount of this soil was thoroly air dried, sieved and mixed. Ten pounds were placed in each of eighty glazed pots, seventy-two of which were given the following treatment: Half, or thirty-six pots received an application of 22.68 grams ground oats straw, and the other half received an equivalent amount of ground red clover hay. This application (22.68 grams) was equivalent to a five-ton application of this material per acre. The ground material was thoroly incorporated in the soil, which was packed firmly in the pots. The pots used were glazed on the inside and made tight so there was no loss by leaching, neither was there any drainage provided.

METHODS OF INOCULATION.

The inoculum used was the dextrose solution described above. 1500 c c were placed in each of six 2 L. flasks, inoculated with the organism desired and incubated for seven days. Microscopic examinations were made at the end of the incubation period to insure vigorous growth and the purity of the culture. 150 cc of the solution was used as the inoculum for each pot. This was poured over the surface of the dry soil and washed into it by the addition of sufficient water to bring the moisture content up to the optimum, in this case 25%. The pots were then weighed, covered with a cloth, and allowed to remain undisturbed for three days, in order to permit the moisture to become thoroly distributed thruout the soil. The pots were then arranged in the following manner and seeded to oats.

Thirty grains of Early Champion oats were planted in each pot at each seeding. They were planted at five points. One in the center of the pot and the other four were arranged between the center and the edge at equal distances apart. Six seeds were planted at each place and when the plants appeared they were thinned out and but one plant left in each place. The discarded plants were allowed to remain and decay on the soil in the pot from which they were drawn.

The length of the growing period was determined by the appearance of the seed-bearing spike. This period varied slightly in each of the series, the first closed in sixty-three days, the second in sixty-nine days, and the third in seventy days after planting.

PLAN OF EXPERIMENT

Pot. No.		Treatment	Inoculation
1— 3	Fallow	Oats straw	A. chroococcum (HCM)
2— 4	Cropped	Oats straw	A. chroococcum (HCM)
5— 7	Fallow	Clover hay	A. chroococcum (HCM)
6— 8	Cropped	Clover hay	A. chroococcum (HCM)
9—11	Fallow	Oats straw	A. chroococcum
10—12	Cropped	Oats straw	A. chroococcum
13—15	Fallow	Clover hay	A. chroococcum
14—16	Cropped	Clover hay	A. chroococcum
17—19	Fallow	Oats straw	A. beijerinckii
18—20	Cropped	Oats straw	A. beijerinckii
21—23	Fallow	Clover hay	A. beijerinckii
22—24	Cropped	Clover hay	A. beijerinckii
25—27	Fallow	Oats straw	A. vinelandii
26—28	Cropped	Oats straw	A. vinelandii
29—31	Fallow	Clover hay	A. vinelandii
30—32	Cropped	Clover hay	A. vinelandii
33—35	Fallow	Oats straw	26 D.
34—36	Cropped	Oats straw	26 D.
37—39	Fallow	Clover hay	26 D.
38—40	Cropped	Clover hay	26 D.
41—43	Fallow	Oats straw	27 D.
42—44	Cropped	Oats straw	27 D.
45—47	Fallow	Clover hay	27 D.
46—48	Cropped	Clover hay	27 D.
49—51	Fallow	Oats straw	22 D.
50—52	Cropped	Oats straw	22 D.
53—55	Fallow	Clover hay	22 D.
54—56	Cropped	Clover hay	22 D.
57—59	Fallow	Oats straw	4 D.
58—60	Cropped	Oats straw	4 D.
61—63	Fallow	Clover hay	4 D.
62—64	Cropped	Clover hay	4 D.
65—67	Fallow	Oats straw	Mixed culture
66—68	Cropped	Oats straw	Mixed culture
69—71	Fallow	Clover hay	Mixed culture
70—72	Cropped	Clover hay	Mixed culture
73—74	Fallow	Oats straw	Check
75—76	Cropped	Oats straw	Check
77—78	Fallow	Clover hay	Check
79—80	Cropped	Clover hay	Check

The pots were watered with tap water about every other day and were weighed weekly. The loss in weight was replaced with water in order to keep the moisture content at the optimum. The growth of the plants was carefully noted and recorded by means of photographs at different periods. The harvested plants were dried, weighed, and the total nitrogen content determined by the Kjeldahl method.

At the close of each series of experiments the soils were removed from the pots, placed on a sterile oil cloth, thoroly mixed, sampled and returned to the original pot. The sample taken at this time approximated 500 grams dry weight. The pots were seeded again as soon as possible and the experiment continued. During the short period between sampling and reseeding the moisture content was kept at the optimum.

The preliminary analyses, showing the nitrogen content of the original air dried soil, and of the same soil mixed with the ground oats or clover are as follows:

22.68 grs. ground oats straw contained.....0.1416 grs. N. av. 6 dets.
22.68 grs. clover hay contained.....0.4153 grs. N. av. 6 dets.
10 lbs. original soil contained.....2.3494 grs. N. av. 6 dets.
10 lbs. original soil + oats straw contained..2.4910 grs. N. av. 6 dets.
10 lbs. original soil + clover hay contained..2.7647 grs. N. av. 6 dets.

ACTION OF DENITRIFYING BACTERIA.

Some of the plants were very much stunted in their growth and an experiment was conducted to determine whether this was due to action by the denitrifying organisms. Samples weighing about six or eight grams were drawn from near the center of each pot by means of a sterile corkborer and placed immediately in sterile tubes. Sterile water was added and a soil suspension made from which inoculations were made into Giltay's denitrifying solution. The solution was incubated three weeks and the amount of nitrate nitrogen as well as the total nitrogen determined, the first by the aluminum reduction method of Potter and Snyder, and the second by the official method. The aluminum reduction was carried out by aeration, thus leaving the original solution available for analysis for total nitrogen. The results given in table VIII show that the denitrifying organisms were not the limiting factor in the growth of plants. Only the five soils in pots Nos. 29, 45, 62, 64 and 66 show any great loss in nitrogen and some of the pots show an actual gain in total nitrogen content. This gain is particularly noticeable in the soils inoculated with *A. beijerinckii,* No. 26 and in the check pots.

TABLE VIII—THE ACTIVITIES OF THE DENITRIFYING BACTERIA IN THE SOILS THREE WEEKS AFTER THE START OF THE EXPERIMENT

Pot	Nitrate N. mgs.	N. mgs.	Total N. mgs.	Check	Amt. denitrified
1	0.70	lost		8.23	
2	1.05	6.02	7.07	8.23	1.16
3	0.98			8.23	
4	0.91	5.60	6.51	8.23	1.72
5	0.90	4.20	5.10	8.23	3.13
6	0.56	8.12	8.68	8.23	*
7	0.70	7.84	8.54	8.23	*
8	0.79	8.14	8.93	8.23	*
9	0.70	7.07	7.77	8.23	0.46
10	0.42	7.07	7.49	8.23	0.74
11	0.70			8.23	
12	0.84	8.96	9.80	8.23	*
13	0.86	6.16	7.02	8.23	1.21
14	0.65	7.56	8.21	8.23	0.02
15	0.56			8.23	

TABLE VIII—Continued.

Pot	Nitrate N. mgs.	N. mgs.	Total N. mgs.	Check	Amt. denitrified
16	1.71	6.16	7.87	8.23	0.36
17	0.56	7.56	8.12	8.23	0.11
18	0.96	7.70	8.66	8.23	:
19	0.42	8.61	9.03	8.23	:
20	0.84	8.40	9.24	8.23	:
21	0.84	7.91	8.75	8.23	:
22	0.56	9.38	9.94	8.23	:
23	0.70			8.23	
24	0.78	7.21	7.98	8.23	0.25
25	0.86	6.44	7.30	8.23	0.93
26	0.06	7.42	7.48	8.23	0.75
27	0.58	8.04	8.62	8.23	:
28	0.87	7.28	8.15	8.23	0.08
29	0.56	5.04	5.60	8.23	2.63
30		6.30		8.23	
31	1.54	5.32	6.86	8.23	1.37
32	0.43	9.10	9.53	8.23	:
33	0.70			8.23	
34	0.56	8.54	9.10	8.23	:
35	0.98	7.56	8.54	8.23	:
36	0.79	8.68	9.47	8.23	:
37	0.86	7.00	7.86	8.23	0.37
38	0.90	6.44	7.34	8.23	0.89
39	1.54	6.58	8.12	8.23	0.11
40	0.63			8.23	
41	0.63	7.56	8.19	8.23	0.04
42	0.56	4.90	5.46	8.23	2.77
43	0.70			8.23	
44	0.98	6.86	7.84	8.23	0.39
45	0.77	2.80	3.57	8.23	4.66
46	1.05	6.30	7.35	8.23	0.88
47	0.77	6.16	7.35	8.23	0.88
48	0.56	6.02	6.58	8.23	1.65
49	0.94	7.28	8.32	8.23	:
50	0.91			8.23	
51	0.81	7.77	8.58	8.23	:
52	0.50	7.70	8.20	8.23	0.03
53	0.77	8.12	8.89	8.23	:
54	0.49	7.56	8.05	8.23	0.18
55	0.86	6.44	7.30	8.23	0.83
56	1.00	6.09	7.09	8.23	1.14
57	0.86	5.46	6.32	8.23	1.81
58	0.86	7.70	8.56	8.23	:
59	0.86	5.60	6.46	8.23	1.77
60	1.33			8.23	
61	0.91			8.23	
62	0.78	1.96	2.74	8.23	5.49
63	1.26	5.60	6.86	8.23	1.37
64	1.31	4.76	6.07	8.23	2.16
65	1.26	7.28	8.54	8.23	:
66	0.98	1.54	2.52	8.23	5.71
67	0.14	7.28	7.42	8.23	0.81
68	0.70	6.16	6.86	8.23	1.37
69	0.56	6.72	7.28	8.23	0.95
70	0.84	4.62	5.46	8.23	2.77
71	0.49	8.40	8.89	8.23	:
72	0.67	7.98	8.65	8.23	:
73	0.87	6.02	6.89	8.23	1.34
74	1.05	7.70	8.75	8.23	:
75	0.65	9.80	10.43	8.23	:
76	0.51	8.12	8.63	8.23	:

*No denitrification is shown by :

PRELIMINARY TESTS FOR NITROGEN FIXATION.

To discover the action of the bacteria in the inoculated soils samples were taken from the fallow pots four weeks after the start of the experiment and their total nitrogen content determined. Table IX shows a gain in the nitrogen content over the original soil and the check soils but the actual gain due to the action of the bacteria introduced was very slight. Organisms 22, 4 and the mixed cultures showed no gain whatever, and the others showed only a slight gain in those soils to which clover had been

TABLE IX—THE ACTIVITY OF THE BACTERIA IN THE INOCULATED FALLOW SOILS AFTER FOUR WEEKS.

Grams of Nitrogen Per 10 Pounds of Soil.

Pot	Grams N. 1st	Grams N. 2nd	Grams N. Av.	Orig. nitrogen content of soil	Nitrogen increase in soils	N. in checks and that in oats and clover hay	Gain due to bact. activities
1	3.0643	lost	3.0643	2.4910	0.5533	0.7288	----------
2	3.0643	3.2281	3.1462	2.4910	0.5552	0.7288	----------
5	3.8942	3.8942	3.8942	2.7647	1.1295	1.0025	0.1270
7	3.8304	3.9581	3.8942	2.7647	1.1295	1.0025	0.1270
9	3.0643	3.2281	3.1462	2.4910	0.6552	0.7288	----------
11	3.2281	3.2281	3.2281	2.4910	0.7371	0.7288	0.0083
13	4.3411	4.0996	4.2203	2.7647	1.4556	1.0025	0.4531
15	3.9581	4.0857	4.0219	2.7647	1.2572	1.0025	0.2547
17	3.2281	3.2281	3.2281	2.4910	0.7371	0.7288	0.0083
19	3.2281	3.0643	3.1462	2.4910	0.6552	0.7288	----------
21	4.2134	4.0219	4.1176	2.7647	1.3529	1.0025	0.3504
23	4.0219	3.9581	3.9900	2.7647	1.2253	1.0025	0.2228
25	3.2281	3.2558	3.2419	2.4910	0.7509	0.7288	0.0221
27	3.2239	3.2239	3.2239	2.4910	0.7329	0.7288	0.0041
29	3.7665	4.0857	3.9261	2.7647	1.1614	0.0025	0.1589
31	4.2772	4.0219	4.1490	2.7647	1.3843	1.0025	0.3818
33	3.2281	3.2281	3.2281	2.4910	0.7371	0.7288	0.0083
35	3.0313	3.1920	3.1281	2.4910	0.6371	0.7288	----------
37	3.8942	3.8304	3.8623	2.7647	1.0976	0.0025	0.0951
39	3.8942	3.8304	3.8623	2.7647	1.0976	1.0025	0.0951
41	3.2281	3.2558	3.2419	2.4910	0.7509	0.7288	0.0221
43	3.1929	3.2281	3.2100	2.4910	0.7109	0.7288	----------
45	3.9581	3.9581	3.9581	2.7647	1.1934	1.0025	0.1909
47	3.7665	3.8304	3.7984	2.7647	1.0337	1.0025	0.0312
49	2.8409	2.8409	2.8409	2.4910	0.3499	0.7288	----------
51	2.9047	2.7679	2.8363	2.4910	0.3453	0.7288	----------
53	3.7027	3.8942	3.7984	2.7647	1.0337	1.0025	0.0312
55	3.5431	3.6069	3.5750	2.7647	0.8103	1.0025	----------
57	2.8409	2.9047	2.8728	2.4910	0.3818	0.7288	----------
59	2.8089	2.7132	2.7610	2.4910	0.2700	0.7288	----------
61	3.5431	3.5752	3.5590	2,7647	0.7943	1.0025	----------
63	3.5750	3.6069	3.5909	2.7647	0.8262	1.0025	----------
65	2.9047	2.8408	2.8727	2.4910	0.3817	0.7288	----------
67	2.7451	2.7451	2.7451	2.4910	0.2541	0.7288	----------
69	3.2239	3.1920	3.2079	2.7647	0.4432	1.0025	----------
71	3.6069	3.4431	3.5250	2.7647	0.7603	1.0025	----------
Checks							
73	2.5536	2.8089	2.6812	2.3494	0.3318		
74	3.0643	3.0324	3.0483	2.3404	0.6989		
77	2.9366	2.8089	2.8727	2.3494	0.5233		
78	3.0643	3.2239	3.1441	2.3494	0.7947		
Average four checks				0.5872 + N in oats		0.1416=0.7288	
				0.5872 + N in clo.		0.4153=1.0025	

added. This difference may have been due to variation in the rate of decomposition between the clover and the straw.

At the end of the three growing periods the soil in each pot was sampled and the total nitrogen content of both the soil and the entire crop determined. The amount of nitrogen found in the determinations and its relation to the total amount due to the bacterial activities is given in three separate tables, one for each growing period. From these complete tables three condensed tables have been made as follows: For the first growing period, table X, for the second growing period, table XI, for the third growing period, table XII, and a recapitulation table XIII.

TABLE X—THE NITROGEN FIXED BY BACTERIA—FIRST PERIOD

(Condensed from Appendix Table I.)

Duplicate pots	Treatment	Bact. Inoculum Used	Grams N. per 10 Pounds Soil			
			N. found	N. in seed—amt. removed by crop	N. in check and in oats and clover	N. fixed by bact.
1—3	F oats	A. chrooc. (HCM)	3.3499	3.3833	3.6232	
2—4	C oats	A. chrooc. (HCM)	2.9883	2.9870	3.7309	
5—7	F clover	A. chrooc. (HCM)	4.3058	4.3488	3.8963	0.4525
6—8	C clover	A. chrooc. (HCM)	4.1808	4.2684	4.0046	0.2638
9—11	F oats	A. chrooc.	3.2210	3.2532	3.6232	
10—12	C oats	A. chrooc.	3.5380	3.5808	3.7309	
13—15	F clover	A. chrooc.	4.5726	4.6183	3.8963	0.7220
14—16	C clover	A. chrooc.	4.4928	4.5787	4.0046	0.5741
17—19	F oats	A. beyer.	5.1701	5.2218	3.6232	1.5986
18—20	C oats	A. beyer.	5.2867	5.2824	3.7309	1.5515
21—23	F clover	A. beyer.	4.4394	4.4827	3.8963	0.5864
22—24	C clover	A. beyer.	4.5801	4.6613	4.0046	0.6567
25—27	F oats	A. vine.	3.9267	3.9659	3.6232	0.3427
26—28	C oats	A. vine.	5.1120	5.1794	3.7309	1.4485
29—31	F clover	A. vine.	2.4572	2.4817	3.8963	
30—32	C clover	A. vine.	4.2388	4.3457	4.0046	0.3411
33—35	F oats	No. 26	3.4887	3.5235	3.6232	
34—36	C oats	No. 26	3.5170	3.5133	3.7309	
37—39	F clover	No. 26	4.4823	4.5271	3.8963	0.6308
38—40	C clover	No. 26	4.5722	4.6833	4.0046	0.6787
41—43	F oats	No. 27	3.6458	3.6822	3.6232	0.0590
42—44	C oats	No. 27	3.5720	3.6217	3.7309	
45—47	F clover	No. 27	4.5023	4.5473	3.8963	0.6510
46—48	C clover	No. 27	4.3114	4.4195	4.0046	0.4149
49—51	F oats	No. 22	3.4101	3.4442	3.6232	
50—52	C oats	No. 22	3.5323	3.5519	3.7309	
53—55	F clover	No. 22	4.4001	4.4441	3.8963	0.5478
54—56	C clover	No. 22	4.3114	4.3866	4.0046	0.3820
57—59	F oats	No. 4	3.6065	3.6425	3.6232	0.0193
58—60	C oats	No. 4	3.5006	3.5323	3.7309	
61—63	F clover	No. 4	4.4630	4.5076	3.8963	0.6113
62—64	C clover	No. 4	4.3023	4.3965	4.0046	0.3919
65—67	F oats	Mixed culture	3.3865	3.4203	3.6232	
66—68	C oats	Mixed culture	3.6038	3.6021	3.7309	
69—71	F clover	Mixed culture	4.3787	4.4123	3.8963	0.5160
70—72	C clover	Mixed culture	4.4720	4.5794	4.0046	0.5748
73—74	F. nothing	Check	3.5942	3.6301		
75—76	C. nothing	Check	3.5085	3.5689		
77—78	F nothing	Check	3.3002	3.3332		
79—80	C. nothing	Check	3.5403	3.6097		

FIRST GROWING PERIOD.

The determinations for this period are shown in appendix table I and in condensed table X. As indicated by table IX, there was a steady increase in the total amount of nitrogen fixed in all the soils. This increase is still more marked if the last columns of tables IX and X are compared. The bacteria were increasingly active in fixing the free atmospheric nitrogen and in practically every case the total amount fixed due to the bacterial solution was more than doubled during the latter five weeks of this series.

These activities may be divided into two classes, as the bacteria were more markedly affected by the presence of clover hay or of oats straw. In the first class *A. chroococcum, A. chroococcum (HCM)*, No. 26, No. 22 and the Mixed Culture stood out prominently. None of these four organisms showed any fixation due to the presence of the decaying oats straw, but they did show appreciable gains due to the presence of the clover hay. The presence of the oats straw had apparently either inhibited the activities of the organisms or increased the activities of the other forms that are incapable of fixing nitrogen for their own use and have utilized that fixed by the inoculating organisms. Organisms 4 and 27 showed a decided stimulation due to the clover hay and were able to utilize the oats straw as a source of energy.

A beijerincki and *A. vinelandii* were more markedly affected by the presence of oats straw. The stimulation of the activities of the former due to the presence of the decaying clover was parallel to that of the other organisms, and in addition the presence of the decaying oats straw stimulated its nitrogen fixing powers to over 250% of that of any other organism in the series with the single exception of *A. vinelandii*. On the other hand, *A. vinelandii*, while showing a marked stimulation due to the presence of the oats, also showed that the clover hay affected its activities much the same as the oats straw affected the other organisms, that is, the presence of the decaying clover hay in the cropped soils, decreased its nitrogen-fixing power, and in the fallow soils, completely inhibited it.

SECOND GROWING PERIOD.

The results for this period are shown in appendix table II, the more important parts of which are repeated in condensed table XI. In comparison with the first growing period the results for the second period are decidedly lower thruout the second series. Not only are the total amounts of nitrogen found lower, but also the total amount of dry matter produced in the crop, indicating a possible direct relation between bacterial action and crop yields. These low results are explained by the fact that this series as

grown during the hottest part of the summer, the pots being planted in the latter part of June and harvested during the earlier part of August. The results confirm those given in Table X except that in this series the only organism stimulated by the presence of the decaying oats straw was organism 27 in the cropped pots. Each of the inoculated organisms showed a direct stimulation due to the presence of the clover hay.

The organisms may be divided into two classes according as their activities are stimulated or retarded by the presence of

TABLE XI—THE NITROGEN FIXED BY BACTERIA—SECOND PERIOD.
(Condensed from appendix Table 2.)

Duplicate Pots	Treatment	Bacterial inoculum used.	Grams Nitrogen per 10 lbs. soil.			
			Nitrogen found.	Nitrogen in seed + amount removed by the growing crop.	Nitrogen in check and in oats and clover.	Nitrogen fixed by bacteria.
1— 3	F. oats	A. chroococcum (HCM)	2.7340	3.0128	3.3370	
2— 4	C. oats	A. chroococcum (HCM)	2.8218	3.0916	3.1866	
5— 7	F. clover	A. chroococcum (HCM)	3.3205	3.6592	3.6107	0.0485
6— 8	C. clover	A. chroococcum (HCM)	3.3123	3.6402	3.4603	0.1799
9—11	F. oats	A. chroococcum	2.6867	2.9607	3.3370	
10—12	C. oats	A. chroococcum	2.5479	2.7947	3.1866	
13—15	F. clover	A. chroococcum	3.5242	3.8836	3.6107	0.2729
14—16	C. clover	A. chroococcum	3.1813	3.4901	3.4603	0.0298
17—19	F. oats	A. Beijerinckii	2.7155	2.9924	3.3370	
18—20	C. oats	A. Beijerinckii	2.6134	2.8626	3.1866	
21—23	F. clover	A. Beijerinckii	3.3343	3.6743	3.6107	0.0636
22—24	C. clover	A. Beijerinckii	3.2681	3.5857	3.4603	0.1254
25—27	F. oats	A. vinelandii	2.8235	3.1115	3.3370	
26—28	C. oats	A. vinelandii	2.6936	2.9763	3.1866	
29—31	F. clover	A. vinelandii	3.3205	3.6591	3.6107	0.0384
30—32	C. clover	A. vinelandii	3.4143	3.6322	3.4603	0.1719
33—35	F. oats	No. 26	2.7547	3.0356	3.3370	
34—36	C. oats	No. 26	2.8912	3.1730	3.1866	
37—39	F. clover	No. 26	3.4921	3.8492	3.6107	0.2385
38—40	C. clover	No. 26	3.0648	3.3747	3.4603	
41—43	F. oats	No. 27	2.7015	2.9770	3.3370	
42—44	C. oats	No. 27	3.0503	3.3429	3.1866	0.1563
45—47	F. clover	No. 27	3.4487	3.8004	3.6107	0.1897
46—48	C. clover	No. 27	3.5089	3.8613	3.4603	0.4010
49—51	F. oats	No. 22	2.7083	2.9845	3.3370	
50—52	C. oats	No. 22	2.8984	3.1700	3.1866	
53—55	F. clover	No. 22	3.3977	3.7442	3.6107	0.1335
54—56	C. clover	No. 22	3.2667	3.5925	3.4603	0.1322
57—59	F. oats	No. 4	2.6146	2.8812	3.3370	
58—60	C. oats	No. 4	2.8246	3.1262	3.1866	
61—63	F. clover	No. 4	3.3908	3.7465	3.6107	0.1358
62—64	C. clover	No. 4	3.1740	3.4913	3.4603	0.0310
65—67	F. oats	Mixed culture	2.5210	2.7781	3.3370	
66—68	C. oats	Mixed culture	2.6935	2.9310	3.1866	
69—71	F. clover	Mixed culture	3.2396	3.6031	3.6107	
70—72	C. clover	Mixed culture	3.3852	3.7159	3.4603	0.2556
73—74	F. nothing	Check	2.8235	3.1115		
75—76	C. nothing	Check	2.7518	3.0537		
77—78	F. nothing	Check	2.9758	3.2793		
79—80	C. nothing	Check	2.7736	3.0364		

growing plants such as clover. In the first class are included *A. chroococcum (HCM), A. beijerinckii, A. vinelandii,* No. 27 and the mixed cultures. The first three organisms have had their nitrogen-fixing powers stimulated by the presence of the plants in practically the same ratio and have fixed similar amounts in both the fallow and cropped soil. *A. beijerinckii* showed the highest fixation of any of the eight for this series. The mixed cultures showed no fixation whatever in the fallow soils, but quite an appreciable amount in the cropped soils. No. 26, on the contrary, fixed an appreciable amount of nitrogen in the fallow soils but none at all in the presence of the growing oats plants. *A. chroococcum* and No. 4 showed the same stimulation under practically the same conditions, namely, that they possess a greater nitrogen-fixing power in the presence of decaying clover if no crop is grown upon the soil, while No. 22 was apparently neither stimulated nor retarded by either fallow or cropped conditions, but was affected by the presence of the decaying oats straw.

THIRD GROWING PERIOD.

The results for this period are shown in appendix table III and condensed table XII. The total nitrogen content of the soil according to tables IX, X and XI, increased steadily through-

TABLE XII—THE NITROGEN FIXATION BY BACTERIA—THIRD PERIOD

(Condensed from Appendix Table III).

Pots	Treatment	Bacterial Inoculum Used	Grams Nitrogen per 10 Pounds Soil				
			N. found	N. in seed + amt. removed by crop	N. in check and in oats and clover	N. fixed by bacteria	Av. N. fixed by bacteria
1	F oats	*A. chrooc.* (HCM)	2.3626	2.8648	2.8502	0.0146	
3	F oats	*A. chrooc.* (HCM)	2.7342	3.3165	2.8502	0.4663	0.2405
2	C oats	*A. chrooc.* (HCM)	2.3643	2,9647	3.3284		
4	C oats	*A. chrooc.* (HCM)	2.1930	2.6754	3.3284		
5	F clover	*A. chrooc.* (HCM)	2.9255	3.5486	3.1239	0.4247	
7	F clover	*A. chrooc.* (HCM)	2.8274	3.4296	3.1239	0.3057	0.3652
6	C clover	*A. chrooc.* (HCM)	3.2852	4.0691	3.6021	0.4670	
8	C clover	*A. chrooc.* (HCM)	2.8688	3.6142	3.6021	0.0121	0.2395
9	F oats	*A. Chroococcum*	2.7489	3.3344	2.8502	0.4842	
11	F oats	*A. Chroococcum*	2.3103	2.8024	2.8502		0.2421
10	C oats	*A. Chroococcum*	2.3652	2.9149	3.3284		
12	C oats	*A. Chroococcum*	2.3785	2.9537	3.3284		
13	F clover	*A. Chroococcum*	2.8928	3.5089	3.1239	0.3850	
15	F clover	*A. Chroococcum*	3.1546	3.8265	3.1239	0.7026	0.5438
14	C clover	*A. Chroococcum*	2.8489	3.5649	3.6021		
16	C Clover	*A. Chroococcum*	2.7501	3.4326	3.6021		
17	F oats	*A. beijerinckii*	2.5789	3.1279	2.8502	0.2777	
19	F oats	*A. beijerinckii*	2.4805	3.0088	2.8502	0.1586	0.2182
18	C oats	*A. beijerinckii*	2.2943	2.8469	3.3284		
20	C oats	*A. beijerinckii*	2.0737	2.5764	3.3284		
21	F clover	*A. beijerinckii*	3.5203	4.2705	3.1239	1.1466	
23	F clover	*A. beijerinckii*	3.0368	3.6836	3.1239	0.5597	0.8532

TABLE XII—Continued

Pots	Treatment	Bacterial Inoculum Used	Grams Nitrogen Per 10 Pounds Soil				
			N. found	N. in seed amt. removed by crop.	N. in check and in oats and clover	N. fixed by bacteria	Av. N. fixed by bacteria
22	C clover	*A. beijerinckii*	2.7429	3.4256	3.6021		
24	C clover	*A. beijerinckii*	2.7694	3.4584	3.6021		
25	F oats	*A. vinelandii*	2.5431	3.0847	2.8502	0.2345	
27	F oats	*A. vinelandii*	2.4478	2.9691	2.8502	0.1189	0.1767
26	C oats	*A. vinelandii*	2.3917	2.9781	3.3284		
28	C oats	*A. vinelandii*	2.3491	2.9494	3.3284		
29	F clover	*A. vinelandii*	3.0172	3.6598	3.1239	0.5359	
31	F clover	*A. vinelandii*	3.0711	3.7252	3.1239	0.6013	0.5686
30	C clover	*A. vinelandii*	2.7429	3.4284	3.6021		
32	C clover	*A. vinelandii*	2.7363	3.3959	3.6021		
33	F oats	No. 26	2.4609	2.9850	2.8502	0.1348	
35	F oats	No. 26	2.3823	2.8887	2.8502	0.0385	0.0862
34	C oats	No. 26	2.4848	3.0673	3.3284		
36	C oats	No. 26	2.3321	2.9746	3.3284		
37	F clover	No. 26	2.9895	3.6162	3.1239	0.4923	
39	F clover	No. 26	2.0164	2.4459	3.1239		0.2462
38	C clover	No. 26	2.7749	3.4477	3.6021		
40	C clover	No. 26	2.8877	3.5888	3.6021		
41	F oats	No. 27	2.4478	2.9691	2.8502	0.1189	
43	F Oats	No. 27	2.4871	3.0169	2.8502	0.1667	0.1428
42	C oats	No. 27	2.4583	3.0736	3.3284		
44	C oats	No. 27	2.4917	3.1595	3.3284		
45	F clover	No. 27	2.8143	3.4137	3.1239	0.2898	
47	F clover	No. 27	3.0564	3.7074	3.1239	0.5835	0.4366
46	C clover	No. 27	3.0181	3.7421	3.6021	0.1400	
48	C clover	No. 27	2.8621	3.5807	3.6021		0.0700
49	F oats	No. 22	2.4961	3.0278	2.8502	0.1776	
51	F oats	No. 22	2.2186	2.6911	2.8502		0.0888
50	C oats	No. 22	2.5580	3.1751	3.3284		
52	C oats	No. 22	2.4451	3.0758	3.3284		
53	F clover	No. 22	2.7230	3.3029	3.1239	0.1790	
55	F clover	No. 22	2.8667	3.4773	3.1239	0.3534	0.2662
54	C clover	No. 22	2.7893	3.5060	3.6021		
56	C clover	No. 22	2.6236	3.2953	3.6021		
57	F oats	No. 4	2.3430	2.8321	2.8502		
59	F oats	No. 4	2.4094	2.9126	2.8502	0.0624	0.0312
58	C oats	No. 4	2.3170	2.8086	3.3284		
60	C oats	No. 4	2.6037	3.3103	3.3284		
61	F clover	No. 4	3.0173	3.6599	3.1239	0.5360	
63	F clover	No. 4	2.8208	3.4216	3.1239	0.2977	0.4168
62	C clover	No. 4	2.8877	3.6172	3.6021	0.0151	
64	C Clover	No. 4	2.8224	3.5098	3.6021		0.0076
65	F oats	Mixed culture	2.3496	2.8501	2.8502		
67	F oats	Mixed culture	2.3234	2.8183	2.8502		
66	C oats	Mixed culture	2.4381	3.0435	3.3284		
68	C oats	Mixed culture	2.3586	2.8937	3.3284		
69	F clover	Mixed culture	3.0761	3.7313	3.1239	0.6074	
71	F clover	Mixed culture	2.7831	3.3759	3.1239	0.2520	0.4297
70	C clover	Mixed culture	2.8366	3.5867	3.6021		
72	C clover	Mixed culture	3.0306	3.7664	3.6021	0.1643	0.0822
73	F nothing	Check	2.3216	2.8161	*	*	*
74	F. nothing	Check	2.0944	2.5405	*	*	*
77	F nothing	Check	2.1072	2.5563	*	*	*
78	F nothing	Check	2.4085	2.9215	*	*	*
75	C nothing	Check	2.7760	3.3649	†	†	†
76	C nothing	Check	2.5375	3.1041	†	†	†
79	C nothing	Check	2.4911	3.1522	†	†	†
80	C nothing	Check	2.4901	3.1362	†	†	†

*Average four fallow checks 2.7086.
†Average four cropped checks 3.1868.

out the first growing period, declined somewhat during the second, and according to table XII, there was a pronounced tendency to increase again during the third period. The crop response of this last period of growth confirms the results of the determinations, the amount of dry matter produced being practically midway between the production of the first and second growing periods. Figs. 1-6, which show the growth of oats in representative pots for the three periods, show that the first crop when ready to harvest was in the majority of cases leafy and heavy and showed a decided tendency to lodge; the second crop in the same stage of growth was somewhat dwarfed in appearance and with no indication of leafiness or weakness of stem; the third crop, while not as heavy as the first, showed all of its characteristics except that as a whole the production was more uniform and did not show the variation in the total dry weight of the harvested crop. The bacterial activities, which are plotted in the tables shown in fig. 7, varied in the same proportion as the crop response of the treated soils, being practically parallel with the production of the dried weight of the crop. The activities increased during the first growing period, declined thruout the second, but increased again during the third. The discussion of the third and last period of growth will be a combination of the activities of the inoculated bacteria as discussed in the first and second growing periods.

The last column in table XII indicates that each inoculated bacterial culture acted without exception in the same general manner instead of showing the expected variations. All of the inoculated bacteria fixed greater amounts of nitrogen in the soils to which clover hay was added as organic matter than in soils that were treated with the same amount of oats straw, and the growing crop on these soils reduced the nitrogen-fixing power of each and every one of these organisms. The activities of any one of the eight organisms used during the third period of growth would be an accurate measure for the activities of any of the others, a fact not even indicated in the other periods of growth.

Conclusion: Table XIII, recapitulating tables X, XI and XII, shows that inoculation, especially in fallow soils to which clover hay or oats straw was added, is not only possible but practical. The amounts of nitrogen shown in these tables are the actual amounts fixed by the organisms in ten pounds of soil and if these amounts are calculated on a 2,000,000 pound acre basis, the result is distinctly profitable. With proper soil conditions the greenhouse experiments can be duplicated in the field.

All of the organisms have shown an appreciable fixation of nitrogen but A. beijerinckii and A. vinelandii have been decidedly the most active. This finding confirms the suggestion of

TABLE XIII—NITROGEN CONTENT OF SOILS AT END OF EACH GROWING PERIOD AND GAIN DUE TO THE INOCULATED BACTERIA. (RESULTS EXPRESSED IN GRAMS PER 10 POUNDS OF SOIL.)

Pots	Treatment	Bacterial inoculum used	Original Nitrogen content of soil + oats or clover	Nitrogen found — that in seed + amount removed by growing crop			Nitrogen fixed by bacteria Average of duplicate pots		
				First Period	Second Period	Third Period	First Period	Second Period	Third Period
1— 3	F. oats	A. chroococcum (HCM)	2.4910	3.3833	3.0128	3.0906			0.2405
2— 4	C. oats	A. chroococcum (HCM)	2.4910	2.9870	3.0916	2.8200			
5— 7	F. clover	A. chroococcum (HCM)	2.7647	4.3488	3.6592	3.4891	0.4525	0.0485	0.3652
6— 8	C. clover	A. chroococcum (HCM)	2.7647	4.2684	3.6402	3.8416	0.2638	0.1799	0.2395
9—11	F. oats	A. chroococcum	2.4910	3.2532	2.9607	3.0684			0.2421
10—12	C. oats	A. chroococcum	2.4910	3.5808	2.7947	2.9343			
13—15	F. clover	A. chroococcum	2.7647	4.6183	3.8836	3.6677	0.7220	0.2729	0.5438
14—16	C. clover	A. chroococcum	2.7647	4.5787	3.4901	3.4987	0.5741	0.0298	
17—19	F. oats	A. beijerinckii	2.4910	5.2218	2.9924	3.0683	1.5986		0.2182
18—20	C. oats	A. beijerinckii	2.4910	5.2842	2.8626	2.6616	1.5515		
21—23	F. clover	A. beijerinckii	2.7647	4.4827	3.6743	3.8779	0.5864	0.0536	0.8532
22—24	C. clover	A. beijerinckii	2.7647	4.6613	3.5857	3.4420	0.6567	0.1254	
25—27	F. oats	A. vinelandii	2.4910	3.9659	3.1155	3.0269	0.3427		0.1767
26—28	C. oats	A. vinelandii	2.4910	5.1794	2.9763	2.9637	1.4485		
29—31	F. clover	A. vinelandii	2.7647	2.4817	3.6591	3.6925		0.0384	0.5686
30—32	C. clover	A. vinelandii	2.7647	4.3457	3.6322	3.4172	0.3411	0.1719	
33—35	F. oats	No. 26	2.4910	3.5235	3.0356	2.9368			0.0862
34—36	C. oats	No. 26	2.4910	3.5133	3.1730	3.0209			
37—39	F. clover	No. 26	2.7647	4.5271	3.8492	3.0310	0.6308	0.2385	0.2472
38—40	C. clover	No. 26	2.7647	4.6833	3.3747	3.5332	0.6787		
41—43	F. oats	No. 27	2.4910	3.6822	2.9770	2.9930	0.0590		0.1428
42—44	C. oats	No. 27	2.4910	3.6217	3.3429	3.1165		0.1563	
45—47	F. clover	No. 27	2.7647	4.5473	3.8004	3.5605	0.6510	0.1897	0.4366
46—48	C. clover	No. 27	2.7647	4.4195	3.8613	3.6614	0.4149	0.4010	0.0700
49—51	F. oats	No. 22	2.4910	3.4442	2.9845	2.8594			0.0888
50—52	C. oats	No. 22	2.4910	3.5519	3.1700	3.1254			
53—55	F. clover	No. 22	2.7647	4.4441	3.7442	3.3901	0.5478	0.1335	0.2662
54—56	C. clover	No. 22	2.7647	4.3866	3.5925	3.4006	0.3820	0.1322	
57—59	F. oats	No. 4	2.4910	3.6425	2.8812	2.8723	0.0193		0.0312
58—60	C. oats	No. 4	2.4910	3.3323	3.1262	3.0594			
61—63	F. clover	No. 4	2.7647	4.5076	3.7465	3.5407	0.6113	0.1378	0.4168
62—64	C. clover	No. 4	2.7647	4.3965	3.4913	3.5635	0.3919	0.0310	0.6076
65—67	F. oats	Mixed culture	2.4910	3.4203	2.7781	2.8342			
66—68	C. oats	Mixed culture	2.4910	3.6021	2.9310	2.9736			
69—71	F. clover	Mixed culture	2.7647	4.4123	3.6031	3.5536	0.5160		0.4297
70—72	C. clover	Mixed culture	2.7647	4.5794	3.7159	3.6765	0.5748	0.2556	0.0822
73—74	F. nothing	Check	2.3494	3.6301	3.1115	2.6783			
75—76	C. nothing	Check	2.3494	3.5689	3.0537	3.2395			
77—78	F. nothing	Check	2.3494	3.3332	3.2793	2.7389			
79—80	C. nothing	Check	2.3494	3.6097	3.0364	3.1442			

FIG. 1. Oats at end of first growing period, immediately before harvest; in pots 2, 6, 10, 14, 18, 22 26, 30, 24, 38

FIG. 2. Oats at end of first growing period, immediately before harvest; pots 44, 48, 52, 56, 60, 64, 68, 72, 76, 80

FIG 3. Oats at end of second growing period, immediately before harvest; pots 2, 6, 10, 14, 18, 22, 26 30, 34, 38

FIG. 4. Oats at end of second growing period, immediately before narvest; pots 42, 46, 50, 54, 58, 62, 66, 70, 75, 79

IG. 5. Oats at end of third growing period, mmediately before harvest; pots 4, 8, 12, 16, 20 24, 28, 32, 36, 40

FIG. 6. Oats at end of third growing period, immediately before harvest; pots 44 48, 52, 56 60 64, 68, 72, 76, 80

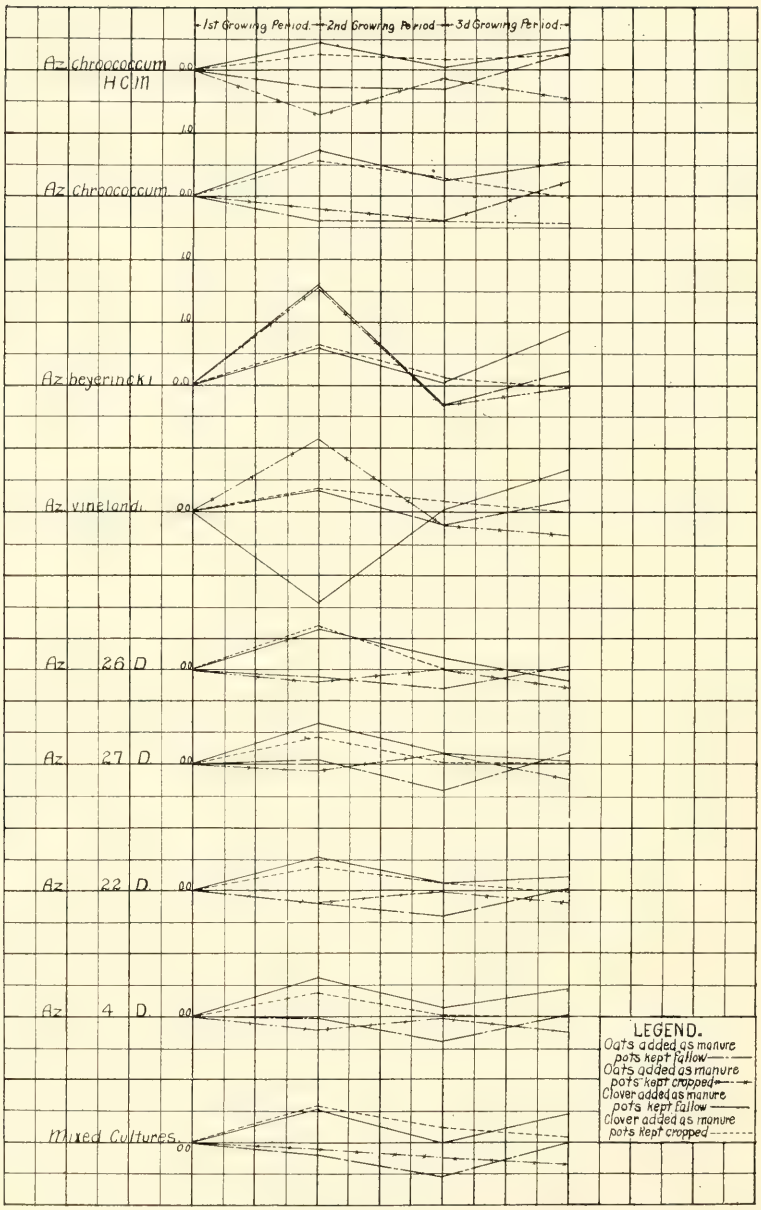

FIG. 7. This graph shows the variation in bacterial activity in the different growing periods

Lipman and Brown (41) by proving definitely that these organisms are capable of being profitably inoculated into field soils, provided that organic matter, carrying a sufficient amount of nitrogen as a stimulus, is supplied.

Summary.

1. When three crops of oats were grown continuously on this soil the nitrogen content of the soil increased during the first cropping period, decreased during the second, and increased slightly again during the third.
2. The nitrogen-fixing powers of the bacteria and the crop response were parallel with the total nitrogen content of the soil.
3. The nitrogen-fixing powers of some types of azotobacter and other large celled organisms of the same general character, were stimulated to a greater extent by the presence of decaying clover hay than of decaying oats straw.
4. The nitrogen-fixing powers of *A. beijerinckii* and *A. vinelandii* were stimulated to a greater extent by decaying oats straw than by clover hay, especially during the earlier stages of decomposition.
5. The nitrogen-fixing powers of the azotobacter and other large celled organisms of the same general type eventually became greater in fallow than in cropped soils.
6. The non-symbiotic nitrogen-fixing organisms of the azotobacter group were all eventually influenced in their activities in the same manner and by the same materials.
7. Soils *may* be profitably inoculated by azotobacter and other large celled organisms of the same type, the best effects being secured in this work by an inoculation with *A. beijerinckii* or *A. vinelandii.*
8. The conditions necessary for the greatest fixation are: Good environmental factors such as tillage, drainage, etc.; the presence of a rapidly decaying organic matter carrying a small nitrogen content, and freedom from growing plants.

ACID EXTRACT, AMINO, NON-PROTEIN AND POLYPEPTID NITROGEN CONTENT OF THE POT SOILS.

Introduction: The nitrogen of the soil is found in many complex combinations, in the determination of which the Bureau of Soils has isolated a large number of nitrogenous compounds and many different forms have been discovered. In investigating methods for the determination of amino acids and nitrates in a limed and unlimed soil, both with and without heavy applications of manures, Potter and Snyder (53) have found that they could accurately measure the amino nitrogen by a modification of the method devised by Kober and Sugiura (32). They discovered

no tendency for the amino acid to accumulate under the conditions of the experiment. Accordingly in the present investigation determinations were made of the acid extract, non-protein, amino, and polypeptid nitrogen of some of the soils inoculated with the azotobacter cultures used in the greenhouse experiments, in order to prove this point and also to discover if the bacterial action had any effect on the accumulation or disappearance of these nitrogenous forms.

Soils used: Only the three soils inoculated with *A. chroococcum, A. beijerinckii* and *A. vinelandii* were analyzed.

METHODS.

Acid extract: Place 166 gr. of air dried soil on a wetted double filter paper in a Buchner funnel and extract with 600 c. c. of a 1% HCl solution using gentle suction. Keep the soil barely covered with the solution and when extracted, wash with 200 to 300 c. c. of pure distilled water. Dry as quickly as possible, and determine the nitrogen content of the filtrate by the official salicylic acid method.

Alkali extract: The non-protein, amino, and polypeptid nitrogen determinations are based on the amounts extracted by a 1.5% NaOH solution. Shake 150 gr. of the air dried acid extracted soil with 600 c. c. of the NaOH solution and centrifuge to a clear solution. At least 210 c. c. of the clear solution must be obtained.

Non-protein nitrogen: Pipette off 25 c. c. of the alkali extract, neutralize with a sulphuric acid solution and add sufficient trichloracetic acid to make a 2.5% solution. To do this use 4.3 c. c., of a 1 3/10 N. H_2SO_4 solution and 0.75 c. c. of a saturated trichloracetic solution. This method precipitates the proteins which are removed by filtering. Pipette 10 c. c. of the clear filtrate into large test tubes, add a couple glass beads, 2 drops of a 5% $CuSO_4$ solution, 1 c. c. C. P. H_2SO_4, and approximately 1 gr. C. P. potassium sulfate. Digest and distil as in the regular Kjeldahl method determining the ammonia colorimetrically.

Amino acid nitrogen: Pipette 80 c. c. of the alkali extract into 100 c. c. measuring flasks, neutralize with strong HCl until neutral to litmus, add 7 c. c. saturated lead acetate solution, fill the flask to the mark with concentrated NH_4OH and shake vigorouly. Allow to settle for a few minutes then pass through double filter, using gentle suction and obtain at least 80 c. c. of the filtrate. Measure off 75 c. c. of this filtrate, add 25 c. c. saturated $Ba(OH)_2$ and phenolphthalein as indicator and distill over steam bath under reduced pressure until there remains a volume of about 25 or 30 c. c. It is important that the reaction of the solution throughout this distillation should be at all times

alkaline. Discard the distillate, wash residue into 100 c. c. graduate, cool, make up to 75 c. c., filter quickly to remove all carbonates, pipette 50 c. c. into 100 c. c. measuring flasks, make approximately neutral with N/10 HCl and add 40 c. c. of buffer solution, stopper tightly and keep in cool place, if possible, on ice. (The buffer is made by dissolving 0.2 gr. molecules of boric acid in water, adding 100 c. c. of CO_2 free N/10 NaOH solution and making up to 1000 c. c. with pure CO_2 free water. Three volumes of this mixed with one volume of O/1/N HCl makes the desired solution.)

Use pure water as cold as possible to prepare fresh the following solution: Place 10-20% copper chloride solution in 20-30 volumes cold water, add a few drops phenolphthalein and a saturated solution $Ba(OH)_2$ until the purple color just forms. Centrifuge, decant off the clear liquid, wash with cold water and recentrifuge, repeating until there is no pink color formed by the addition of phenolphthalein in the wash water. Suspend the copper hydroxide in about 100 c. c. cold water and add approximately 1 c. c. to the cool flasks, shake vigorously, make up to the mark, and allow to warm up to the room temperature. Filter through No. 589 blue ribbon filter, pipette off 50 c. c. of the filtrate and determine the copper complex present as shown below as a measure of the amino nitrogen. Pipette off 40 c. c. of the filtrate for the determination of the polypeptid nitrogen.

Polypeptid nitrogen: Hydrolize the polypeptids into amino acids by adding approximately 5 c. c. concentrated H_2SO_4 to the 40 c. c. and placing under a steam pressure of 8-10 pounds for 10-12 hours. Remove the excess acid with a saturated solution $Ba(OH)_2$ keeping the solution slightly alkaline to phenolphthalein, filter and wash with carbonated water at least three times. Evaporate the filtrate to about 35 or 40 c. c., place in 100 c. c. measuring flasks, neutralize with N/10 HCl, add 40 c. c. buffer solution, 1 c. c. of the copper hydroxide solution in the cold water as for the amino determinations and determine the copper present in the same manner.

Copper determination: Place the beakers containing the 50 c. c. on the hot plate, heat to boiling and neutralize with dilute HNO_3. Boil down to about one-half and add bromine water until a decided bromine color appears, evaporate to about 10-15 c. c., add 20-30 c. c. pure water and a little more bromine water and evaporate down again to 10-15 c. c. Cool, add 2-3 c. c. glacial acetic acid, a few crystals potassium iodide, a few drops of starch solution and titrate immediately with .001/N sodium thio-sulfate until the blue color disappears. Each c. c. of the .001/N thio-sulfate solution is the equivalent of 0.000028 gr, amino acid nitrogen.

Preliminary determinations: In addition to the work on the soils, an unsuccessful attempt was made to determine the amount of non-protein and amino acid nitrogen fixed by the bacteria inoculated into the dextrose solution used in the other experiments. 250 c. c. of the dextrose solution was inoculated with the organisms indicated in Table 14 and incubated three weeks at room temperature. Enough c. p. sodium hydroxide was added directly to the solution to make 1.5% and the determination carried out in the above manner. A slight trace was the greatest amount found.

This table shows a decided increase in the soils under field conditions over the same soils in the dry state, the greatest increase taking place during the earlier periods of growth. The results of these determinations are grouped in three tables, each showing the amount of the different nitrogenous forms found at the end of each growing period.

Discussion of results: A comparison of the results given in Tables XVI, XVII and XVIII, shows that there was a definite variation of the nitrogenous forms with the length of the time of cropping. In almost every case the amount extracted by the acid varied with the length of time that the soil had been cropped, growing smaller and smaller, and the amino and polypeptid nitrogen gave similar results. The amount of these nitrogenous

TABLE XIV—AMINO ACID AND NON-PROTEIN NITROGEN FIXED BY THE PURE CULTURES IN SOLUTION.

Flask	Inoculum	Non-protein N.	Amino Acid N.
1	A. chroococcum (HCM)	------	trace
2	A. chroococcum (HCM)	------	------
3	A. chroococcum (HCM)	------	------
4	No. 26	------	trace
5	No. 26	------	------
6	No. 26	trace	------
7	A. chroococcum (HCM) and No. 26	trace	trace
8	A. chroococcum (HCM) and No. 26	------	trace
9	A. chroococcum (HCM) and No. 26	------	------

TABLE XV—THE AMOUNT OF DIFFERENT NITROGENOUS FORMS IN THE SOIL AT THE BEGINNING OF THE EXPERIMENTS, ALSO THE SAME SOIL PLUS THE EQUIVALENT OF FIVE TONS GROUND OATS, STRAW OR GROUND CLOVER HAY ADDED TO THE SAMPLE. DETERMINATIONS BASED ON THE AMOUNT IN 25 GR. OF THE SAMPLE AND RESULTS EXPRESSED IN MG. NITROGEN AND IN PER CENT OF THE TOTAL NITROGEN.

Soil	Original Nitrogen content	Acid ex-N, mg. found	Per cent	Non-protein N. mg. found	Per cent	Amino acid N. mg. found	Per cent	Poly-peptid N. mg. found	Per cent
Original	12.95	1.1242	8.7	2.2275	17.2	0.6840	0.7	0.2100	1.6
Original + oats	13.75	1.1666	8.5	2.3390	17.0	0.1025	0.7	0.2550	1.8
Original + red clover hay	15.40	1.4424	9.3	2.3475	15.2	0.1050	0.7	0.2625	1.7

TABLE XVI—AMOUNTS OF THE DIFFERENT FORMS OF NITROGEN IN THE CROPPED AND FALLOW INOCULATED SOILS AT THE END OF THE FIRST PERIOD OF GROWTH. RESULTS EXPRESSED IN MG. NITROGEN FOUND AND IN PER CENT OF THE TOTAL NITROGEN CONTENT, BASED ON 25 GR. SAMPLE.

Pots	Total N. content 25 gr.	Acid extract mg. found	Per cent	Non-protein mg. found	Per cent	Amino acid mg. found	Per cent	Poly-peptid mg. found	Per cent
9—11	14.35	2.0242	14.1+	3.1675	22.0	0.1260	0.8+	0.4725	3.2+
10—12	15.60	1.9181	12.3—	2.6650	17.0+	0.1505	0.9+	0.2975	1.9
13—15	20.39	2.2151	10.8+	2.3690	11.6+	0.1400	0.7—	0.4550	2.2+
14—16	19.81	1.9818	10.0+	3.0650	15.4+	0.1085	0.5+	0.2000	1.0+
17—19	23.03	2.2939	9.9+	3.5000	15.2+	0.1015	0.4+	0.2625	1.1+
18—20	23.09	1.1121	4.7+	2.8500	12.3+	0.1680	0.7+	0.2100	0.9+
21—23	19.77	2.3424	11.8+	2.7175	13.8—	0.2310	1.1+	0.4200	2.1+
22—24	20.19	0.9848	4.8+	2.7325	13.5+	0.1400	0.7—	0.3500	1.7+
25—27	17.50	2.4030	13.7+	2.6950	15.4	0.1820	1.0+	0.4375	2.5
26—28	22.54	1.9878	8.8+	2.7425	12.1+	0.2110	0.9+	0.3150	1.4—
29—31	20.94	2.3848	11.4—	2.5150	12.0+	0.1750	0.8+	0.2100	1.0
30—32	18.69	0.7515	4.1—	2.5875	13.8+	0.0980	0.5+	0.3200	1.7+

TABLE XVII—AMOUNTS OF THE DIFFERENT FORMS OF NITROGEN IN THE FALLOW AND CROPPED INOCULATED SOILS AT THE END OF THE SECOND GROWING PERIOD. RESULTS EXPRESSED IN MG. N. FOUND AND IN PER CENT OF THE TOTAL N. CONTENT BASED ON 25 GRAM SAMPLE.

Pots	Total N. content 25 gr.	Acid extract mg. found	Per cent	Non-protein mg. found	Per cent	Amino acid N. mg. found	Per cent	Polypeptid N. mg. found	Per cent
9—11	16.45	1.6515	10.0	1.8375	11.1+	0.0700	0.4+	0.2800	1.7+
10—12	15.44	lost	----------	3.4150	22.1+	0.1110	0.7+	0.3675	2.4—
13—15	21.48	2.1896	10.2	2.0700	9.6+	0.1260	0.6—	0.1600	0.7+
14—16	19.32	0.7257	3.7+	2.9925	15.5—	0.1400	0.7+	0.1225	0.6+
17—19	16.62	0.9212	5.5+	2.6825	16.1+	0.4750	2.8+	0.1750	1.1—
18—20	15.84	0.7257	4.6—	3.4075	21.5+	0.0201	0.1+	0.3500	2.2+
21—23	19.74	1.7895	9.1—	3.5000	17.8—	0.1190	0.6—	0.2100	1.0+
22—24	19.78	0.7500	3.8—	3.3350	16.8+	0.0630	0.3+	0.2800	1.4+
25—27	17.28	1.4727	8.5+	2.6370	15.3—	0.2660	1.5+	0.2100	1.2+
26—28	16.32	0.6030	3.7—	2.6000	15.9+	0.0630	0.4—	0.2975	1.8+
29—31	20.31	2.1363	10.5+	2.8350	13.9+	0.0910	0.4+	0.2800	1.3+
30—32	20.66	1.1000	5.4+	3.4900	16.9—	0.0700	0.3+	0.2275	1.1+

TABLE XVIII—AMOUNTS OF THE DIFFERENT FORMS OF NITROGEN IN THE FALLOW AND CROPPED INOCULATED SOILS AT THE END OF THE THIRD AND LAST GROWING PERIOD. RESULTS BASED ON 25 GRAM SAMPLE, EXPRESSED IN MG. N. FOUND AND IN PER CENT OF TOTAL N. CONTENT.

Pots	Total N. content 25 gr.	Acid extract mg. found	Per cent	Non-protein N. mg. found	Per cent	Amino acid N. mg. found	Per cent	Polypeptid mg. found	Per cent
9—11	16.93	1.2833	7.5+	2.1475	12.7—	0.0420	0.2+	0.2275	1.3+
10—12	15.65	0.5773	3.7—	3.2850	20.9+	0.0490	0.5+	0.2100	1.3+
13—15	20.23	0.7485	3.7—	2.3675	11.2+	0.0420	0.2+	0.2450	1.2+
14—16	18.19	0.6306	3.4+	3.3005	18.1+	0.0910	0.4+	0.2100	1.1+
17—19	16.90	1.1773	6.9+	2.7025	15.9+	0.0210	0.1+	0.2625	1.6—
18—20	14.94	0.5560	3.7+	3.6825	24.7—	0.0630	0.4+	0.1400	0.9+
21—23	22.03	1.7727	8.1—	2.8560	12.9+	0.0420	0.2—	0.2800	1.3—
22—24	18.21	0.7000	3.8+	3.0300	16.5+	0.3240	1.8—	0.1750	0.9+
25—27	16.71	1.5060	9.0+	2.6925	16.1+	0.0350	0.2+	0.5950	3.6—
26—28	15.66	0.7212	4.6+	3.6350	23.2+	0.0420	0.3—	0.3150	2.0+
29—31	20.36	2.2060	10.8+	2.7400	13.4+	0.0560	0.4+	0.7075	3.4+
30—32	18.09	0.5929	3.3—	3.4500	11.3—	0.0770	0.4+	0.2800	1.5+

compounds became smaller, as decomposition of the organic matter proceeded, at a slightly faster rate than the total nitrogen content of the soil became depleted. The non-protein nitrogen also varied considerably, altho not in the marked degree shown by the other forms. Neither the oats straw or the red clover hay, added as manures to the pots, showed any effect on the forms of nitrogen determined, further than the small amount shown in the preliminary determinations. If there was a difference in the soil under field conditions it evidently was too small to be measured by these methods. It is entirely possible that the amounts of these complex nitrogenous compounds are rapidly changing into other forms and that the per cent they bear to the total nitrogen content remains somewhat constant, varying only with the amount of organic matter present in the beginning, then as decomposition proceeds and the more complex combinations are broken up, this percentage relation becomes smaller and smaller until it reaches a constant.

Once decomposition had begun in the soil there was absolutely no tendency for the more complex nitrogenous forms to accumulate under conditions approximating those in the field. Instead of an accumulation there was a steady reduction. How closely this reduction is coupled with the decay of the organic matter and what would be the final equilibrium between the total nitrogen content and the nitrogenous compounds are questions for further study.

Summary.

1. The acid extracted, non-protein, amino and polypeptid nitrogen changed into other forms with the advance of decomposition much faster than the total nitrogen contents of the soils in question decreased.

2. Oats straw and clover hay added as manures at the rate of five tons per acre had little effect in influencing this change.

3. The amounts of non-protein and amino acid nitrogen fixed by bacterial cultures in solution were negligible.

4. Bacterial inoculation had apparently no effect on the amounts of non-protein, amino or polypeptid nitrogen in the soil.

5. There was no tendency for the above forms of nitrogen to accumulate in the soil under conditions approximating those in the field.

Acknowledgments: I wish to express my thanks to Dr. P. E. Brown for his help and suggestions thruout this work and to Dr. R. S. Potter for his suggestions in the determinations of the complex forms of nitrogen.

APPENDIX TABLE I.

Inoculum Used and Pot No.	Treatment	Kind of Crop Grown	Actual Grams N. Found in Pot Soils, Calculated on Basis of 4491 gr. in Fallow and 4536 gr. in cropped.—Pot Determinations: Orig.	Dup.	Av.	Less amt. N. added in seed	Total gr. N. in pot	Dry wt. of crop grams	Total N. content of crop grams	Total N. content of pot at time of adding N. in seed+N. removed by crop	N. content on basis of gr. vol. of soil, 4,536 gr.	Less check to which has been added the N. content of the dried oats or clover but few that was added to other pots	Grams N. gain due entirely to action of the inoculum
Az. chrooc. (HCM)													
1- 3	F	Oats	3.6369	3.3630	3.3499	.....	3.5499						
2- 4	O	Oats	2.7658	3.2079	2.9883	0.0136	2.9747	0.8350	0.0123	3.3499	3.3833	3.6222	.....
5- 7	F	Clover	4.2744	4.3373	4.3058	.....	4.3058			2.9870	2.9870	3.7350	.....
6- 8	C	Clover	4.1671	4.2547	4.1809	.....	4.1673	7.1150	0.1011	4.3058	4.3488	3.8963	0.1453
Az. chrooccum										4.2584	4.2584	4.6046	0.2638
9-11	F	Oats	2.8601	3.5820	3.2210	.....	3.2210						
10-12	O	Oats	3.4450	3.6310	3.5380	0.0136	3.5244	3.7400	0.0564	3.2210	3.2332	3.6222	.....
13-15	F	Clover	4.5663	4.5798	4.5726	.....	4.5726			3.5868	3.5868	3.7309	.....
14-16	C	Clover	4.4779	4.5687	4.4923	0.0136	4.4787	6.7600	0.0995	4.5726	4.6183	3.8963	0.7129
A. beijerinckii										4.4787	4.4787	4.6046	0.5711
17-19	F	Oats	5.1230	5.2173	5.1701	.....	5.1701						
18-20	O	Oats	5.1392	5.2343	5.2367	0.0136	5.2751	0.8250	0.0093	5.1701	5.2218	3.6222	1.5595
21-23	F	Clover	4.5702	4.5687	4.4394	.....	4.4394			5.2324	5.2324	3.7309	1.5015
22-24	C	Clover	4.5881	4.5722	4.5861	0.0136	4.5665	7.0100	0.0948	4.4394	4.4827	3.8963	0.5864
A. vinelandii										4.6613	4.6613	4.6046	0.6597
25-27	F	Oats	3.9601	3.9573	3.9267	.....	3.9267						
26-28	O	Oats	3.1755	3.0485	3.1120	0.0136	3.0984	5.0050	0.0810	3.9267	3.9639	3.6222	0.3427
29-31	F	Clover	4.2651	4.4494	4.4572	.....	4.4572			5.1794	5.1794	3.7309	1.4485
30-32	C	Clover	4.2865	4.1012	4.2383	0.0136	4.2222	8.2950	0.1205	4.4572	4.4817	3.8963	0.5854
Az. 26D										4.3457	4.3457	4.6046	0.3411
33-35	F	Oats	3.4887	3.4887	3.4857	.....	3.4857						
34-36	O	Oats	3.4927	3.5463	3.5170	0.0136	3.5034	0.4900	0.0099	3.4857	3.5235	3.6222	.....
37-39	F	Clover	4.4703	4.4944	4.4823	.....	4.4823			3.5133	3.5133	3.7309	.....
38-40	C	Clover	4.6040	4.5405	4.6722	0.0136	4.5586	7.9650	0.1247	4.4823	4.5271	3.8963	0.6308
Az. 27D										4.6633	4.6633	4.6046	0.6587
41-43	F	Oats	3.5830	3.7087	3.6458	.....	3.6458						
42-44	O	Oats	3.5562	3.5679	3.5720	0.0136	3.5584	3.7500	0.0633	3.6458	3.6842	3.6222	0.0590
43-45	F	Clover	4.4473	4.5573	4.5023	.....	4.5023			3.6217	3.6217	3.7309	.....
44-46	C	Clover	4.3500	4.2729	4.3114	0.0136	4.2978	9.6450	0.1217	4.5023	4.5476	3.8963	0.6610
Az. 28D										4.4195	4.4195	4.6046	0.4149
49-51	F	Oats	3.3630	3.4573	3.4101	.....	3.4101						
50-52	O	Oats	3.4768	3.5879	3.5323	0.0136	3.5187	2.1850	0.0322	3.4101	3.4112	3.6222	.....
53-55	F	Clover	4.4316	4.3687	4.4001	.....	4.4001			3.5519	3.5519	3.7309	.....
54-56	C	Clover	4.2720	4.3500	4.3114	0.0136	4.2978	6.7850	0.0888	4.4001	4.4141	3.8963	0.5247
Az. 4D										4.3806	4.3806	4.6046	0.3220
57-59	F	Oats	3.6144	3.5987	3.6065	.....	3.6065						
58-60	O	Oats	3.4627	3.5685	3.5906	0.0136	3.4870	3.8650	0.0453	3.6065	3.6425	3.6222	0.0203
61-63	F	Clover	4.4316	4.4944	4.4630	.....	4.4630			3.5323	3.5323	3.7309	.....
62-64	C	Clover	4.2230	4.3817	4.3023	0.0136	4.2887	7.1400	0.1073	4.4630	4.5076	3.8963	0.6113
Mixed cultures										4.3065	4.3065	4.6046	0.3919
65-67	F	Oats	3.3315	3.4415	3.3865	.....	3.3865						
66-68	O	Oats	3.5362	3.6514	3.6038	0.0136	3.5902	1.0450	0.0119	3.3865	3.4203	3.6222	.....
69-71	F	Clover	4.4567	.....	4.4567	.....	4.4567			3.6021	3.6021	3.7309	.....
70-72	C	Clover	4.4185	4.5405	4.4720	0.0136	4.4584	9.5300	0.1210	4.4567	4.4123	3.8963	0.5166
Check										4.5791	4.5791	4.6046	0.5748
73-74	F	Nothing	3.6369	3.6515	3.5942	.....	3.5942					Average	
74-76	O	Nothing	3.5562	3.4609	3.5685	0.0136	3.4949	5.5650	0.0740	3.5942	3.6301	3.4816	.....
77-78	F	Nothing	3.3002	3.3002	3.3002	.....	3.3002			3.5639	3.5639	3.7309	.....
79-80	O	Nothing	3.4609	3.6197	3.5403	0.0136	3.5267	6.7700	0.0830	3.3002	3.3329	.....	.....
										3.6097	3.6097	.....	.....

APPENDIX TABLE II.

Inoculum Used and Pot No.	Treatment	Kind of Crop Grown	Actual Grams N. Found in Pot Soils, Calculated on Basis 4116 gr. in Fallow, 4161 gr. in Cropped.—Pots Determination: Orig.	Dup.	Av.	Amt. added in seed	Total gr. N. in pot	Dry wt. of crop grams	Total N. content of crop grams	Total N. in pot when sample was taken—amt. N. added in seed+amt. N. removed by the growing crop	Column 11 expressed on basis of orig. volume of soil, 4656 gr.	Less check to which has been added the N. content of the dried oats or clover but few added to other pots	Grams N. gain due entirely to action of the inoculum
Az. chrooc. (HCM)													
1- 3	F	Oats	2.7371	2.7309	2.7340	.....	2.7340				2.7340	3.0128	3.3370
2- 4	O	Oats	2.8837	2.7000	2.8218	0.0136	2.8082	1.4995	0.0252	2.8864	3.0916	3.1866	.....
5- 7	F	Clover	3.3854	3.2207	3.3205	.....	3.3205			2.8082	2.8082	3.6107	0.4859
6- 8	C	Clover	3.1070	3.2177	3.1123	0.0136	3.2987	2.1740	0.0410	3.3397	3.6402	3.4603	0.1750
Az. chrooccum													
9-11	F	Oats	2.6795	2.6939	2.6867	.....	2.6867				2.6867	2.9607	3.3370
10-12	O	Oats	2.5183	2.5771	2.5479	0.0136	2.5343	2.7877	0.0257	2.5610	2.7947	3.1866	.....
13-15	F	Clover	3.5191	3.5274	3.5242	.....	3.5242			3.5242	3.8836	3.6107	0.2729
14-16	C	Clover	3.1888	3.1740	3.1813	0.0136	3.1677	1.9253	0.0343	3.2020	3.4901	3.4603	0.0298
A. beijerinckii													
17-19	F	Oats	2.7227	2.7083	2.7155	.....	2.7155				2.7155	2.9924	3.3370
18-20	O	Oats	2.6353	2.5916	2.6134	0.0136	2.5998	1.8895	0.0265	2.6263	2.8926	3.1866	.....
21-23	F	Clover	3.2269	3.4318	3.3343	.....	3.3343			3.3343	3.6743	3.6107	0.0536
22-24	C	Clover	3.2322	3.3051	3.2681	0.0136	3.2545	1.8340	0.0452	3.2897	3.5857	3.4603	0.1254
Az. vinelandii													
25-27	F	Oats	2.8379	2.8091	2.8235	.....	2.8235				2.8235	3.1115	3.3370
26-28	O	Oats	2.6936	2.6936	2.6936	0.0136	2.6800	2.0178	0.0506	2.7306	2.9763	3.1866	.....
29-31	F	Clover	3.3277	3.3133	3.3205	.....	3.3205			3.3205	3.6591	3.6107	0.0334
30-32	C	Clover	3.4070	3.4216	3.4143	0.0136	3.4007	3.1785	0.0629	3.4636	3.6322	3.4603	0.1719
Az. 26D													
33-35	F	Oats	2.7517	2.7577	2.7547	.....	2.7547				2.7547	3.0356	3.3370
34-36	O	Oats	2.8537	2.9286	2.8912	0.0136	2.8776	2.2545	0.0344	2.9120	3.1730	3.1866	.....
37-39	F	Clover	3.5443	3.4460	3.4921	.....	3.4921			3.4921	3.8402	3.6107	0.2385
38-40	C	Clover	3.0867	3.0459	3.0648	0.0136	3.0512	2.2800	0.0449	3.0961	3.3747	3.4603	.....
Az. 27D													
41-43	F	Oats	2.6795	2.8235	2.7015	.....	2.7015				2.7015	2.9770	3.3370
42-44	O	Oats	2.9411	3.1595	3.0503	0.0136	3.0367	1.6869	0.0302	3.0669	3.3129	3.1866	0.1563
43-45	F	Clover	3.4374	3.4100	3.4487	.....	3.4487			3.4487	3.8004	3.6107	0.1897
44-46	C	Clover	3.5089	3.5089	3.5089	0.0136	3.4953	1.8225	0.0472	3.5425	3.8613	3.4603	0.4010
Az. 28D													
49-51	F	Oats	2.7227	2.6939	2.7083	.....	2.7083				2.7083	2.9845	3.3370
50-52	O	Oats	2.8633	2.6236	2.7384	0.0136	2.8748	1.8625	0.0335	2.9083	3.1700	3.1866	.....
53-55	F	Clover	3.3977	3.3977	3.3977	.....	3.3977			3.3977	3.7442	3.6107	0.1335
54-56	C	Clover	3.3051	3.2333	3.2692	0.0136	3.2331	2.5103	0.0428	3.2959	3.5625	3.4603	0.1322
Az. 4D													
57-59	F	Oats	2.6218	2.6074	2.6146	.....	2.6146				2.6146	2.8812	3.3370
58-60	O	Oats	2.8537	2.7955	2.8246	0.0136	2.8110	0.3305	0.0571	2.8681	3.1262	3.1866	.....
61-63	F	Clover	3.3854	3.4142	3.3998	.....	3.3998			3.3998	3.7465	3.6107	0.1358
62-64	C	Clover	3.1740	3.1740	3.1740	0.0136	3.1604	2.2090	0.0427	3.2031	3.4913	3.4603	0.0310
Mixed cultures													
65-67	F	Oats	2.5066	2.5354	2.5210	.....	2.5210				2.5210	2.7781	3.3370
66-68	O	Oats	2.7081	2.6700	2.6935	0.0136	2.6799	2.2315	0.0422	2.7221	2.9810	3.1866	.....
69-71	F	Clover	3.2696	3.2696	3.2696	.....	3.2696			3.2696	3.6031	3.6107	.....
70-72	C	Clover	3.4216	3.3188	3.3852	0.0136	3.3716	1.5933	0.0375	3.4091	3.7159	3.4463	0.2556
Check												Average	
73-74	F	Nothing	2.8091	2.8379	2.8235	.....	2.8235			2.8235	3.1115	*	*
74-76	C	Nothing	2.7518	2.7518	2.7518	0.0136	2.7382	3.0903	0.0469	2.7851	3.0377	†	†
77-78	F	Nothing	2.9696	2.8839	2.9268	.....	2.9268			2.9268	3.2793	‡	‡
79-80	O	Nothing	2.7955	2.7518	2.7736	0.0136	2.7600	1.0500	0.0257	2.7857	3.0364	‖	‖

*3.1951 + WL Oats N. = 3.3370
†3.0450 + WL Oats N. = 3.1866
‡3.1951 + clover N. 3.6107
‖3.0450 + clover N. = 3.4603

APPENDIX TABLE III.

Inoculum Used and Pot No.	Treatment	Kind of Crop Grown	Actual Grams N. Found in Pot Soils, Calculated on Basis of 3741 gr. in Fallow and 3786 gr. in Cropped Pots Determinations: Orig.	Dup.	Av.	N. added in seed	Total gr. N. in pot	Dry wt. of crop in gr.	Total N. content of crop in gr.	Total N. in pot at amplitude—N. in seed+N. removed by crop.	Total N. expressed on basis of orig. vol. of soil, 4536 gr. mass	Less check to which has been added the N. content of the oats or clover added as manure	Gain due entirely to action of inoculum expressed as gr. N.
Az. chrooc. (HCM)													
1- 3	F	Oats	2.3360	2.3953	2.3629	.....	2.3626				2.3626	2.8648	2.8502	0.0116
2- 4	O	Oats	2.3434	2.3882	2.3643	0.0136	2.3507	3.4020	0.1338	2.3626	2.4063	2.9647	3.0284	.....
5- 7	F	Clover	2.7019	2.6965	2.7342	.....	2.7342			2.3507	2.7342	3.3165	2.8502	0.1065
6- 8	C	Clover	2.1364	2.1996	2.1930	0.0136	2.1794	2.5110	0.0447	2.7342	2.9241	2.6754	3.3284	.....
Az. chrooccum										2.1794	2.1794	2.6754	3.1350	0.1247
9-11	F	Oats	2.8923	2.9583	2.9253	.....	2.9253				2.9253	3.5496	3.1350	0.1247
10-12	O	Oats	3.2694	3.3711	3.3202	0.0136	3.3066	3.6030	0.1109	2.9253	3.2776	3.8826	3.6021	0.4670
13-15	F	Clover	2.8143	2.8405	2.8274	.....	2.8274			3.3066	3.3066	4.0601	3.1350	0.2647
14-16	C	Clover	2.9417	2.7359	2.8553	0.0136	2.8553	3.4310	0.1492	2.8274	2.8274	3.4296	3.6021	0.0121
Az. chrococcum										2.8553	3.0044	3.6142	3.6021	0.0121
17-19	F	Oats	2.7489	2.7489	2.7489	.....	2.7489				2.7489	3.3344	2.8502	0.4842
18-20	O	Oats	2.3321	2.3984	2.3653	0.0136	2.3516	2.5590	0.0715	2.7489	2.4231	2.9149	3.3284	.....
21-23	F	Clover	2.9088	2.8169	2.8163	.....	2.8163			2.3516	2.8163	3.3963	2.8502	0.5660
22-24	C	Clover	2.3586	2.3984	2.3785	0.0136	2.3649	2.7750	0.0604	2.8163	2.4855	2.9957	3.3284	.....
25-27	F	Clover	2.9059	2.8798	2.8928	.....	2.8928			2.3649	2.8928	3.4889	3.1350	0.3560
26-28	C	Clover	2.8763	2.8234	2.8499	0.0136	2.8363	3.7675	0.1251	2.8928	2.9434	3.6449	3.6021	.....
29-31	F	Clover	3.1808	3.1285	3.1546	.....	3.1546			2.8363	3.1546	3.8265	3.1350	0.7029
30-32	C	Clover	2.5971	2.7032	2.7501	0.0136	2.7361	3.3540	0.1175	3.1546	3.2554	3.4326	3.6021	.....
Az. beijerinckii										2.7361	2.7361	3.3191	3.6021	.....
33-35	F	Oats	2.5918	2.5656	2.5787	.....	2.5787				2.5787	3.1279	2.8502	0.2777
34-36	O	Oats	2.2860	2.3227	2.3043	0.0136	2.2907	0.7500	0.0195	2.5787	2.3566	2.8589	3.3284	.....
37-39	F	Clover	2.5132	2.4478	2.4805	.....	2.4805			2.2907	2.4805	3.0088	2.8502	0.1586
38-40	C	Clover	2.6804	2.6670	2.6737	0.0136	2.6601	3.2680	0.0816	2.4805	2.4417	2.9564	3.3284	.....
41-43	F	Clover	2.8763	2.8393	2.8578	.....	2.8578			2.6601	2.8578	3.4705	3.1350	0.1166
42-44	C	Clover	2.7429	2.7429	2.7429	0.0136	2.7293	3.4500	0.1185	2.8578	2.8256	3.4256	3.6021	.....
43-45	F	Clover	2.9976	3.0761	3.0368	.....	3.0368			2.7293	3.0368	3.6878	3.1350	0.5597
44-46	C	Clover	2.7694	2.7694	2.7694	0.0136	2.7553	3.4220	0.1101	3.0368	2.8749	3.4884	3.6021	.....
Az. vinelandii										2.7553				
49-51	F	Oats	2.5132	2.5731	2.5431	.....	2.5431				2.5431	3.0847	2.8502	0.2345
50-52	O	Oats	2.3851	2.3983	2.3917	0.0136	2.3781	0.9075	0.0253	2.5431	2.4756	2.9781	3.3284	.....
53-55	F	Clover	2.4478	2.4478	2.4478	.....	2.4478			2.3781	2.4478	2.9681	2.8502	0.1189
54-56	C	Clover	2.3684	2.3719	2.3701	0.0136	2.3565	3.1830	0.1166	2.4478	2.4517	2.9494	3.3284	.....
57-59	F	Clover	3.0107	3.0237	3.0172	.....	3.0172			2.3565	3.0172	3.6698	3.1239	0.5559
58-60	C	Clover	2.7429	2.7429	2.7429	0.0136	2.7293	3.8030	0.1289	3.0172	2.8382	3.4384	3.6021	.....
61-63	F	Clover	3.0499	3.0923	3.0711	.....	3.0711			2.7293	3.0711	3.7222	3.1239	0.6603
62-64	C	Clover	2.7297	2.7422	2.7360	0.0136	2.7227	3.1575	0.1002	3.0711	2.8229	3.4059	3.6021	.....
Az. 26D										2.7227				
32-34	F	Oats	2.4478	2.4740	2.4609	.....	2.4609				2.4609	2.9850	2.8502	0.1348
33-35	O	Oats	2.4653	2.5044	2.4848	0.0136	2.4712	3.4015	0.0753	2.4609	2.5497	3.0673	3.3284	.....
34-36	F	Clover	2.3682	2.3964	2.3823	.....	2.3823			2.4712	2.3823	2.8867	2.8502	0.0365
35-37	C	Clover	2.3586	2.3056	2.3321	0.0136	2.3185	2.2160	0.0542	2.3823	2.4727	2.9746	3.3284	.....
37-39	F	Clover	2.3976	2.9714	2.9895	.....	2.9895			2.3185	2.9895	3.6162	3.1239	0.1023
38-40	C	Clover	2.7429	2.7969	2.7749	0.0136	2.7613	3.9045	0.1396	2.9895	2.8090	3.4077	3.6021	.....
39-41	F	Clover	2.0102	2.0027	2.0064	.....	2.0064			2.7613	2.0064	2.4459	3.1239	.....
40-42	C	Clover	2.9152	2.8602	2.8877	0.0136	2.8741	2.9730	0.1091	2.0064	2.9832	3.6388	3.6021	.....
Az. 27D										2.8741				
41-43	F	Oats	2.4478	2.4478	2.4478	.....	2.4478				2.4478	2.9691	2.8502	0.1189
42-44	O	Oats	2.4513	2.4653	2.4583	0.0136	2.4447	3.6430	0.1103	2.4478	2.5550	3.0736	3.3284	.....
43-45	F	Clover	2.4871	2.4871	2.4871	.....	2.4871			2.4447	2.4871	3.0160	2.8502	0.1667
44-46	O	Oats	2.5189	2.4646	2.4917	0.0136	2.4781	4.0130	0.1483	2.4871	2.6396	3.1695	3.3284	.....
45-47	F	Clover	2.7881	2.8405	2.8143	.....	2.8143			2.4781	2.8143	3.4142	3.1239	0.2828
46-48	C	Clover	2.8814	2.8548	2.8681	0.0136	2.8545	3.5770	0.1321	2.8143	3.1206	3.7421	3.6021	0.1469
47-49	F	Clover	3.0499	3.0630	3.0564	.....	3.0564			2.8545	3.0564	3.7074	3.1239	0.5834
48-50	C	Clover	2.9019	2.8224	2.8621	0.0136	2.8485	3.8010	0.1230	3.0564	2.9765	3.5857	3.6021	.....
Az. 22D										2.8485				
49-51	F	Oats	2.5494	2.4478	2.4961	.....	2.4961				2.4961	3.0278	2.8502	0.1776
50-52	O	Oats	2.5448	2.5713	2.5580	0.0136	2.5444	2.9430	0.0949	2.4961	2.6393	3.1751	3.3284	.....
51-53	F	Clover	2.2353	2.1951	2.2186	.....	2.2186			2.5444	2.2186	2.6911	2.8502	.....
52-54	C	Clover	2.4116	2.4789	2.4453	0.0136	2.4315	3.7480	0.1253	2.2186	2.5568	3.0768	3.3284	.....
53-55	F	Clover	2.7032	2.7429	2.7230	.....	2.7230			2.4315	2.7230	3.3029	3.1239	0.1790
54-56	C	Clover	2.7837	2.7959	2.7898	0.0136	2.7757	4.0300	0.1387	2.7230	2.9144	3.5069	3.6021	.....
55-57	F	Clover	2.8907	2.8907	2.8907	.....	2.8907			2.7757	2.8907	3.4773	3.1239	0.3531
56-58	C	Clover	2.0767	2.5706	2.6236	0.0136	2.6100	3.5470	0.1222	2.8907	2.7392	3.2953	3.6021	.....
Az. 4D										2.6100				
57-59	F	Oats	2.3169	2.3692	2.3430	.....	2.3430				2.3430	2.8821	2.8502	.....
58-60	O	Oats	2.3000	2.3321	2.3160	0.0136	2.3024	2.3930	0.0426	2.3430	2.3450	2.8686	3.3284	.....
59-61	F	Clover	2.4116	2.4789	2.4453	.....	2.4453			2.3024	2.4453	3.0768	2.8502	0.0654
60-62	O	Oats	2.6164	2.5971	2.6068	0.0136	2.5931	4.4220	0.1616	2.4453	2.7537	3.3103	3.3284	.....
61-63	F	Clover	3.0109	3.0237	3.0173	.....	3.0173			2.5931	3.0173	3.6599	3.1239	0.3560
62-64	C	Clover	2.8002	2.9152	2.8577	0.0136	2.8441	4.3390	0.1327	3.0173	3.0068	3.6172	3.6021	0.0071
63-65	F	Clover	2.8843	2.8374	2.8608	.....	2.8608			2.8441	2.8608	3.4216	3.1239	0.2767
64-66	C	Clover	2.7570	2.8469	2.8324	0.0136	2.8188	3.1685	0.1057	2.8608	2.9175	3.5098	3.6021	.....
Mixed cultures										2.8441				
65-67	F	Oats	2.3431	2.3562	2.3496	.....	2.3496				2.3496	2.8501	2.8502	.....
66-68	O	Oats	2.4619	2.4116	2.4381	0.0136	2.4245	3.4770	0.0935	2.3496	2.5309	3.0635	3.3284	.....
67-69	F	Oats	.....	2.3231	2.3231	.....	2.3231			2.4245	2.3231	2.8182	2.8502	.....
68-70	O	Oats	2.3189	2.3984	2.3586	0.0136	2.3450	2.9405	0.0604	2.3231	2.4654	2.9937	3.3284	.....
69-71	F	Clover	3.0109	3.1023	3.0761	.....	3.0761			2.3450	3.0761	3.7313	3.1239	0.6674
70-72	O	Clover	2.8773	2.7959	2.8366	0.0136	2.8230	4.3355	0.1383	3.0761	2.9815	3.5867	3.6021	.....
71-73	F	Clover	2.7489	2.8274	2.7831	.....	2.7831			2.8230	2.7831	3.3769	3.1239	0.2530
72-74	C	Clover	2.9947	2.9695	3.0306	0.0136	3.0170	3.1908	0.1168	2.7831	2.9180	3.5681	3.6021	0.0340
Check										3.0306				
73-74	F	Nothing	2.3189	2.3244	2.3216	.....	2.3216				2.3216	2.8161	.....	.....
74-76	F	Nothing	2.0944	2.0944	2.0944	.....	2.0944			2.3216	2.0944	2.5495	.....	.....
75-77	O	Nothing	2.7694	2.7827	2.7760	0.0136	2.7624	1.6355	0.0339	2.0944	2.7963	3.3949	.....	.....
76-78	O	Nothing	2.5574	2.5176	2.5375	0.0136	2.5239	2.5740	0.0339	2.7624	2.5803	3.1041	.....	.....
77-79	F	Nothing	2.1074	2.1074	2.1074	.....	2.1074			2.5239	2.1074	2.5563	.....	.....
78-80	F	Nothing	2.4216	2.3954	2.4085	.....	2.4085			2.1074	2.4085	2.9215	.....	.....
79-81	O	Nothing	2.5176	2.4646	2.4911	0.0136	2.4775	3.9540	0.1436	2.4085	2.6211	3.1522	.....	.....
80-82	O	Nothing	2.4513	2.5189	2.4901	0.0136	2.4765	3.5010	0.1304	2.4775	2.6069	3.1361	.....	.....

Av. ck. fallow 2.7086 + N. content of oats ground 0.1416=2.8502
Av. ck. fallow 2.7086 + N. content ground clover 0.4163=3.1239.
Av. ck. cropped 3.1868 + N. content ground oats 0.1416=3.3284.
Av. ck. cropped 3.1868 + N. content ground clover 0.4163=3.6021

BIBLIOGRAPHY

1. ASHBY, S. F.
   1907. Some observations on the assimilation of atmospheric nitrogen by a free-living organism-*Azotobacter chroococcum* of Beijerinck. Journ. Agr. Sci. **2**:35.
2. BEIJERINCK, M. W.
   1901. Nitrogen fixing bacteria. Centbl. f. Bakt. 2 Abt. **7**:561.
3. BEIJERINCK, M. W.
   1908. Fixation of nitrogen by pure culture of azotobacter; distribution of the organism. K. Akad. Wetensch. Amsterdam, Proc. Sect. Sci. **11**:67.
   1908. Abs. in Journ. Chem. Soc. **94**:975.
4. BEIJERINCK, M. W. and VAN DELDEN, A. V.
   1902. The assimilation of free nitrogen by bacteria. Centbl. f. Bakt. 2. Abt. **9**:3.
5. BOLLEY, H. L.
   1900. The duration of bacterial existence and trial environments. Centbl. f. Bakt. 2 Abt. 6:33.
6. BONAZZI, A.
   1915. Cytological studies of *Azotobacter chroococcum*. Journ. Agr. Rsch. **4**:225.
7. BOTTOMLEY, W. B.
   1907. Cereals and bacteria. Country Life, **22**:466.
8. BOTTOMLEY, W. B.
   1910. The fixation of nitrogen by free-living soil bacteria. Proc. Roy. Soc. Ser. B. **82**:627.
   1910. Abs. in Journ. Soc. Chem. Indus. **29**:1171.
9. BOTTOMLEY, W. B. and HALL, A. D.
   1909. Nitrogen-fixing bacteria and non-leguminous plants. Nature **82**:218.
10. CHRISTENSEN, H. R.
    1906. On the occurrence and distribution of *Azotobacter chroococcum* in different soils. Centbl. f. Bakt. **17**:109, 161, 378.
11. CZAPLEWSKI, E.
    1889. Bacteriological methods. Centbl. f. Bakt. Abt. **6**:409.
12. DVORAK, J.
    1912. Studies on nitrogen fixation in soil by micro-organisms. Zeitschr. f. d. Landw. Versuchswes in Oster. Jahrg. **15**:1077.
    1913. Abs. in Centbl. f. Bakt. 2 Abt. **37**:106.
13. EHRENBERG, P.
    1909. The bearing of carbon determinations upon nitrogen fixation in soils. Fühling's Landw. 2tg. **58**:663.
14. FISCHER, H.
    1905. Contribution to the knowledge of the life conditions of nitrogen-collecting bacteria. Centbl. f. Bakt. 2 Abt. **14**:33, 781.
15. FISCHER, H.
    1905. Nitrogen fixing bacteria. Journ. Landw. **53**:61.
16. FISCHER, H.
    1905. Contribution to the knowledge of the life conditions of nitrogen-collecting bacteria. Journ. Landw. **53**:289.
17. FRENDENREICH, E. DE
    1903. Assimilation of atmospheric nitrogen by bacteria. Centbl. f. Bakt. 2 Abt. **10**:514.

18. GERLACH, M. and VOGEL, I.
1902. Nitrogen-gathering bacteria. Centbl. f. Bakt. 2 Abt. **8**:669; **9**:816; **9**:881.
19. GERLACH, M., and VOGEL, I.
1903. Further studies on nitrogen gathering bacteria. Centbl. f. Bakt. 2 Abt. **10**:636.
20. GREAVES, J. E., and ANDERSON, H. P.
1914. The influence of arsenic upon the nitrogen fixing powers of the soil. Centbl. f. Bakt. 2 Abt. **42**:244.
21. HEADDEN, W. P.
1910. The fixation of nitrogen in some Colorado soils. Bull. Colo. Agr. Exp. Sta. 155.
1911. Bull. Colo. Agr. Exp. Sta. 178.
1913. Bull. Colo. Agr. Exp. Sta. 186.
22. HEINZE, B.
1906. Some contributions to the microbiology of soils. Centbl. f. Bakt. 2 Abt. **16**:640.
23. HEINZE, B.
1906. Concerning nitrogen fixation by lower organisms. Landw. Jahr. **35**:889.
24. HOFFMAN, C.
1913. The protein and phosphorus content of azotobacter cells. Centbl. f. Bakt. 2 Abt. **36**:474.
25. HOFFMAN, C., and HAMMER, B. W.
1911. Some factors concerned in the fixation of nitrogen by azotobacter. Res. Bull. Wisc. Agr. Exp. Sta. 12.
26. JACOBITZ, E.
1901. The assimilation of free atmospheric nitrogen. Centb. f. Bakt. 2 Abt. **7**:783, 833, 876.
27. JONES, D. H.
1913. A morphological and cultural study of some azotobacter. Centbl. f. Bakt. 2 Abt. **38**:14.
28. KASERER, H.
1910. On the mineral needs of azotobacter. Ber. d. Deutsch. Bot. Gesell. **28**:208.
29. KELLERMAN, K. F., and SMITH, N. R.
1914. The absence of nitrate formation in cultures of azotobacter. Centbl. f. Bakt. 2 Abt. **40**:479.
30. KOCH, A.
1910. On nitrogen fixation in soils with cellulose as a source of energy. Centbl. f. Bakt. 2 Abt. **27**:1.
1907. Mitt. Deut. Landw. Gesell. **22**:117.
1909. Journ. Landw. **57**:269.
31. KOCH, A., and SEYDEL, S.
1911. The utilization of cellubiose as a source of energy in nitrogen fixation through bacteria. Centbl. f. Bakt. **31**:567.
32. KOBER, P., and SUGIURA, S.
1913. A micro-chemical method for the determination of alpha and beta amino acids and certain derivatives in proteolysis, blood and urine. Journ. Amer. Chem. Soc. **35**:1546.
33. KRAINSKY, A.
1908. *Azotobacter chroococcum* and its action in the soil. Centbl. f. Bakt. 2 Abt. **20**:725.
1910. Centbl. f. Bakt. 2 Abt. **26**:231.
34. KUHN, J.
1901. The assimilation of free nitrogen by soil bacteria without symbiosis with leguminous plants. Fühling's Landw. Ztg. **50**:2.

35. LIPMAN, J. G.
1904. Soil bacteriological studies. Rpt. N. J. Agr. Exper. Sta. **1904**:237.
36. LIPMAN, J. G.
1903. Nitrogen-fixing bacteria. Doctor's thesis, Cornell University.
37. LIPMAN, J. G.
1903. Experiments on the transformation and fixation of nitrogen by bacteria. Rpt. N. J. Agr. Expt. Sta. **1903**:247.
38. LIPMAN, J. G.
1904. Soil bacteriological studies. Rpt. N. J. Agr. Exp. Sta. **1904**:263.
39. LIPMAN, J. G.
1902. Nitrogen-fixing bacteria. Pop. Sc. Monthly. **62**:97.
1903. Rpt. N. J. Agr. Exp. Sta. **1903**:217.
1904. Bull. Bur. Chem. **81**:146.
40. LIPMAN, J. G. and BROWN, P. E.
1911. Laboratory guide in soil bacteriology.
41. LIPMAN, J. G., and BROWN, P. E.
1907. Report of the soil chemist and bacteriologist. Rpt. N. J. Agr. Exp. Sta. **1907**:141.
42. LIPMAN, C. B., and BURGESS, P. S.
1915. Studies on nitrogen fixation and azotobacter forms in soils in foreign countries. Centbl. f. Bakt. 2 Abt. **44**:481.
43. LÖHNIS, F.
1909. The importance of nitrogen fixation in cultivated soil. Fühling's Landw. Ztg. **58**:425.
44. LÖHNIS, F., and HANZAWA, J.
1914. The classification of azotobacter. Centbl. f. Bakt. **42**:1.
45. LÖHNIS, F., and PILLAI, N. K.
1908. Nitrogen-fixing bacteria. Centbl. f. Bakt. 2 Abt. **20**:781.
46. LÖHNIS, F., and PILLAI, N. K.
1907 Nitrogen-fixing bacteria. Centbl. f. Bakt. 2 Abt. **19**:87.
47. LÖHNIS, F., and SMITH, N. R.
1916. Life cycles of bacteria. Journ. Agr. Rsch. **6**:675.
48. LÖHNIS, F., and WESTERMANN, T.
1908. Nitrogen-fixing bacteria. Centbl. f. Bakt. 2 Abt. **22**:234.
49. LUTZ, L.
1905. The morphology and biology of nitrogen-assimilating organisms. Centbl. f. Bakt. **15**:477.
50. MULVANIA, M.
1915. Observations on azotobacter. Science, N. S. **42**:463.
51. OMELIANSKY, W. L. and SSEWEROWA, O. P.
1911. Pigment formation in cultures of *Azotobacter chroococcum* Centbl. f. Bakt. 2 Abt. **29**:643.
52. PETERSON, E. G., and MOHR, E.
1913. Non symbiotic nitrogen fixation by organisms in Utah soils. Centb. f. Bakt. 2 Abt. **38**:494.
53. POTTER, R S., and SNYDER, R. S.
1915. Determination of amino acids and nitrates in soils. Res. Bull. Ia. Agr. Exp. Sta. 24.
54. PRAZMOWSKI, A.
1912. Natural history, morphology and cytology of *Azotobacter chroococcum*. Centbl. f. Bakt. 2 Abt. **33**:292.
55. PRINGSHEIM, H.
1907. On the adaptability of different sources of energy to assimulation of atmospheric nitrogen and the distribution of nitrogen-fixing bacteria in the soil. Centbl. f. Bakt. 2 Abt. **20**:248.
1908. Centbl. f. Bakt. 2 Abt. **26**:222.

56. PRINGSHEIM, H.
1914. Nitrogen assimilation in the presence of nitrates. Centbl. f. Bakt. 2 Abt. **40**:21.
57. REMY, T.
1909. Investigations on the process of nitrogen assimilation in its relation to the soil climate. Centbl. f. Bakt. 2 Abt. **22**:561.
58. REMY, T., and ROSING, G.
1911. The biological stimulation of natural humus. Centbl. f. Bakt. 2 Abt. **30**:349.
59. ROSING, G.
1912. Results of the investigations on nitrogen fixation by *Azotobacter chroococcum*. Centbl. f. Bakt. 2 Abt. **33**:292; 618.
60. STOKLASSA, J.
1908. Contribution to the knowledge of the chemical processes involved in the assimilation of free nitrogen by azotobacter and radiobacter. Centbl. f. Bakt. 2 Abt. **21**:484.
61. STRANAK, J.
1909. The assimilation of atmospheric nitrogen by soil micro-organisms. Centbl. f. Bakt. 2 Abt. **25**:320.
62. THIELE, R.
1906. The utilization of atmospheric nitrogen by micro-organisms. Landw. Vers. Sta. **63**:161.
63. VAGLER, P.
1909. The fixation of atmospheric nitrogen. Centbl. f. Bakt. 2 Abt. **22**:452.
64. VOGEL, J.
1912. Potash requirements by the nitrogen bacteria. Centbl. f. Bakt. 2 Abt. **32**:411.
65. VOGEL, J.
1903. Some recent investigations in nitrogen assimilation by bacteria without symbiosis. Fühling's Landw. Ztg. **52**:178.
1903. Reference Exp. Sta. Rec. **14**:1048.
66. VORHEES, E. B., and LIPMAN, J. G.
1905. Experiments on the accumulation and utilization of atmospheric nitrogen in the soil. Bull. N. J. Agr. Exp. Sta. 180.
67. WILFARTH, H., and WIMMER, G.
1907. The influence of mineral fertilization on the nitrogen fixation by lower organisms in the soil. Landsw. Vers. Sta. **67**:27.
68. WARMBOLD, H.
1906. Investigations on the biology of the nitrogen fixing bacteria. Landw. Jahs. **35**, No. 1-2; 1., **35**, No. 4-6; 63.

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